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REVIEW



Bioactive compounds from agroindustrial byproducts and their applications: From traditional methods to green technologies

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Abstract

The use of agroindustrial byproducts (solids and liquids) with bioactive compounds has developed and grown remarkably. Existing methodologies and some new approaches developed through various research efforts have demonstrated their viability. The bioactive compounds commonly studied in research are phenolic compounds and carotenoids because of their potential as antioxidants and natural additives that can prolong the shelf life of certain foods. Meanwhile, conventional extraction methods are being replaced with green technologies, which offer reduced extraction time and are more environmentally friendly, among other advantages. In addition, protecting and preserving bioactive compounds from environmental conditions by applying microencapsulation and nanoencapsulation methods is crucial. These studies have shown that combining methods yields better results than using them individually. Therefore, it is recommended that unconventional methods and green technologies be integrated into future research. This approach will help determine whether acquiring them is profitable and feasible for the agroindustrial companies responsible for this waste. This review demonstrates the potential use of agricultural byproducts to obtain bioactive compounds (through combined extraction methods for improved efficiency) and incorporate them in different industries, such as food, cosmetics, and aquaculture, thereby creating a favorable socio-environmental impact by reducing soil and water pollution.

Keywords: Agroindustrial wastewater and solids; effluents; extraction; encapsulation; environmental impact.

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1. Introduction

As agribusiness has grown recently, byproducts from fruit and vegetable processing have increased. Despite being rich in scientifically interesting compounds, these byproducts often become waste. This situation generates increased environmental pollution, highlighting the need to apply circular economy principles in production processes. It is necessary to first utilize all byproducts or waste generated during the processing of value-added products and improve their preservation method. This allows for the extraction of bioactive compounds that can be applied in foods to provide functional and nutritional properties for consumers (Figueroa et al., 2021).

Espinosa et al. (2019) have shown that byproducts, such as husks and seeds, are an important source of polyphenols and carotenoids capable of providing antioxidant power and natural color to certain

food products. To obtain, conserve, and protect bioactive compounds from environmental factors, researchers extract and encapsulate them using various techniques or methodologies. The most used extraction methods are conventional, ultrasound-assisted, and microwave extraction. For microencapsulation, spray drying is the most common technique owing to its low cost and high performance compared to other techniques. After application, they are characterized physicochemical and sensorily to evaluate the viability of the final product fortified with bioactive compounds. Studies have repeatedly shown that the attributed improvements are valid and extend the product's useful life (Surco et al., 2020).

This review presents information on recent scientific research regarding extraction and encapsulation methods and their applications across various industries. It summarizes the most used methods,

highlights their key differences, and discusses their characterization after application.

2. Agroindustrial byproducts

Agroindustrial byproducts are residues obtained during the processing of certain raw materials in the agroindustrial sector that have no use in the production chain. In large quantities, they become food waste, often owing to inadequate transport and storage conditions within the agroindustrial production chain (Angulo et al., 2018). However, they are an excellent source of bioactive compounds, such as phenolic compounds (hydroxycinnamic acid, hydroxybenzoic acid, flavanols, and flavonols) and carotenoids (carotenes and xanthophylls), whose consumption prevents or reduces the risk of chronic noncommunicable diseases (Jofre et al., 2020).

The use of agroindustrial byproducts reduces food waste and improves food security and the efficiency of the agrifood system. Foods that generate the highest levels of waste during their production chain include tubers, roots, and oilseed species (25%), followed by fruits and vegetables to a lesser extent (20% – 22%). This waste comprises primary waste (leaves, roots, and stems) and secondary waste (husks and seeds), with the latter containing higher concentrations of bioactive compounds, such as polyphenols and carotenoids (Martínez Rodríguez, 2021).

Studies have shown that fruit and vegetable peels are a crucial source of bioactive compounds. Robles et al. (2020) showed that the concentrations of tannins, flavan-3-ol, and proanthocyanidins in the husk of tejocote (*Crataegus mexicana*) were substantially higher than that of the pulp and seed. This is because sunlight is needed to synthesize flavonoids, which are mainly located in the outer tissues of plants. Coca (2021) stated that pomegranate (*Punica granatum*) peel contains phenolic acids, flavonoids, hydrolyzable tannins, and proanthocyanidins. It primarily comprises punicalagin, the most important bioactive compound (phenolic compound) in this fruit, and possesses antimicrobial properties. Marentes et al. (2019) obtained 165 ± 9 mg cyanidin-3-O-glucoside/L of anthocyanins from the epicarp or shell of gulupa (*Passiflora edulis f. edulis*), demonstrating it to be a byproduct with high potential as a source of anthocyanins, surpassing even purple sweet potato (158 mg anthocyanin/100 g body weight). Similarly, Ku (2021) mentioned that cainito (*Chrysophyllum cainito*) peels (tropical fruit) are a rich source of anthocyanins and other phenolic compounds.

Other common byproducts of this industry are seeds containing phenolic compounds (flavonoids

and anthocyanins) and carotenoids, from which food additives, such as thickening agents, colorings, and flavorings, can be obtained. For example, avocado seeds are high in phenolic compounds, such as caffeic acid, ferulic acid, 3-O-p-cumaroylquinic acid, and isomers of chlorogenic acid, rendering them rich in antioxidants and offering potential in the food industry (Flores et al., 2021). Capillo & Navarro (2019) extracted 456.25 mg EAG/g of polyphenols from ungurahui (*Oenocarpus bataua*) seed using an ultrasonic bath. The most abundant compounds were flavonoids and tannins. Coriat (2022) obtained 523.330 ± 1.95 mg GAE/100 g of camu camu (*Myrciaria dubia*) seeds by optimizing the Soxhlet method, concluding that over 190% of the phenolic compounds of the byproduct can be recovered.

Other agroindustrial residues have also been studied for their use, such as olive oil byproducts (pomace and alpechin), that have been shown to possess a high content of phenolic compounds. Cea (2020) determined 2865.7 mg EAG/kg of polyphenols on a dry pomace basis, which are mostly secoiridoid (54.8%) and flavonoids (23%). In addition, 34909.3 mg EAG/kg of polyphenols on a dry basis of alpechin were characterized, identifying mainly phenolic alcohols, secoiridoids, flavonoids, and verbascosides. Fruit and vegetable leaves are byproducts of agroindustry that are nutrient-rich and high in antioxidants owing to their bioactive compounds. While some are utilized for animal feed, most are wasted due to improper usage. Cervantes-Sierra et al. (2019) determined that the polyphenol content extracted from purple sweet potato leaves was 14.16 mg of gallic acid GAE/g, which surpasses that of green tea leaves (9.40 mg of gallic acid GAE/g) that was also studied in their research. Jordan (2019) stated that soursop (*Annona muricata*) leaves contain phenolic compounds, such as chlorogenic acid, catechin, quercetin, caffeic acid, and gallic acid. Other bioactive compounds include acetogenins (such as anonacin, muricin, montanacin, and solamine) and alkaloids (such as anonain, anonamine, reticulín, and isoboldin). The study demonstrated that the polyphenol content extracted from soursop leaves was 49.03 mg AG/g and the amount of anonacin was 0.05 mg/g.

Another type of agroindustrial waste with a high content of organic components of interest is liquid effluents. These effluents are characterized by high concentrations of solids with organic and pathogenic loads, which leads to the death of aquatic species, the formation of sediment layers, and the development of unpleasant odors. However,

agroindustrial effluents have a considerable content of phenolic compounds that depend on the type of fruit, the degree of ripeness, and the volume of water used during processing (mainly in the washing stage). Effluents from tomato processing, winemaking, and olive trees are rich in flavonoids and carotenoids. **Corredor & Pérez (2018)** have shown that solid agroindustrial byproducts can effectively remove dye from liquid effluents. For example, rice husks have been used to remove 99% of methylene blue dye, 91% of basic red dye, and up to 72.8% of chromium when activated with phosphoric acid and sodium hydroxide. When chemically modified with sulfuric, hydrochloric, and phosphoric acids, sugarcane bagasse and corn stover can remove heavy metals with up to 98.6% effectiveness. Other residues include tangerine peels, banana peels, and coffee residues. Although this area has been studied in recent years, there remains limited information on the benefits of this type of waste to reduce pollutant loads and protect water resources (**Vuksinic, 2019**).

Agroindustry effluents can be transformed into value-added products such as food ingredients, biomaterials (adsorbents for heavy metals, nanomaterials, and building reinforcements), chemicals, fuels, bioenergy, polymers, antioxidants, pharmaceuticals, fertilizers, livestock feed, lipids, and phenols. This transformation helps reduce food waste and the environmental impact. However, the recovery of these valuable compounds depends on the quantity of value-added molecules they possess, and the treatment methods applied (**Bustillos-Sánchez et al., 2022**). Agroindustrial activities, such as animal slaughter, bioethanol production, and the processing of oil palm, cassava, milk, and sugarcane, generate huge amounts of wastewater, mainly used for energy production (methane or hydrogen) because of organic compounds. Although agroindustrial effluents are a considerable source of micronutrients, such as chromium, cobalt, copper, iron, zinc, manganese, molybdenum, nickel, and selenium, they contain substantial amounts of toxic compounds, such as heavy metals, which can bioaccumulate in the food chain (De Carvalho et al., 2022). In cassava processing wastewater, some vitamins such as vitamins B2 and C have also been found; however, their use as food additives and pharmaceutical ingredients must be carefully considered based on a thorough study of their composition, and no examples of pilot or industrial production are known yet. Phenolic compounds have been extracted from liquid effluents derived from vanilla and wine using eutectic and bio solvents (**Cañadas Soler, 2022**).

Furthermore, agroindustrial effluents have been used as a source of nutrients for some fungi and bacteria through fermentation processes to produce bioproducts, such as enzymes, pigments, biosurfactants, antibiotics, and phenolic compounds, for various industrial applications. In addition, this approach offers an interesting alternative for water decontamination. Olive mill wastewater is an agroindustrial waste reported to be an adequate substrate for producing bioproducts via fermentation, specifically submerged fermentation. This process occurs in excess free water and is considered an effective solution for treating liquid waste while obtaining enzymes and compounds with antityrosinase activities via *Pleurotus citrinopileatus*. Other enzymes produced using olive mill wastewater as a substrate are L-asparaginase by *Bacillus aryabhattai* and n-demethylases. L-asparaginase is a key chemotherapeutic agent in acute lymphoblastic leukemia. Palm oil mill effluent is a source of nutrients for lipase production by *Pseudomonas aeruginosa*. Enzyme production by fermentation using agroindustrial liquid effluents as raw material could have enormous biotechnological potential due to its environmental and pharmaceutical applications (**Astudillo et al., 2023**).

3. Types of bioactive compounds in agroindustrial byproducts

Bioactive compounds are phytochemicals that have gained considerable interest recently due to their beneficial properties in foods containing them. They can be classified into phenolic compounds, terpenoids, and thiols. Phenolic compounds comprise anthocyanins, catechins, flavonoids, isoflavones, lignans, and tannins. Terpenoids are divided into carotenoids, capsaicin, and phytosterols. In addition, thiols comprise organosulfur compounds. Although they are not considered nutrients, as they do not provide calories (such as vitamins and minerals), they can have synergistic physiological effects accompanied by other substances (**Urrialde et al., 2022**). The most studied bioactive compounds in food byproducts are polyphenols and carotenoids.

3.1 Phenolic compounds

Aguilar-Méndez et al. (2020) has demonstrated that phenolic compounds were extracted from ungurahui seeds, which were pulverized for quantification, obtaining 452.76 mg EAG/g EUL of polyphenols. Phenolic compounds were also extracted from the peel and pulp of two native potato varieties, with the concentrations compared. The peel was observed to have higher levels than the pulp for both varieties (between 628.89 and

877.67 mg AGE/100 g) using the conventional method. Using ultrasound-assisted extraction, the range obtained was 1770.25 ± 27.13 – 1910.42 mg AGE/100 g. Similar extractions were performed on avocado, cocoa, coconut, and prickly pear shells. The highest phenolic content extracted was from avocado shells with 36.5 ± 0.5 mg GAE/g dry weight, followed by coconut shells with 13.6 ± 0.5 mg GAE/g dry weight. Cocoa and prickly pear shells obtained about 10 ± 0.5 mg GAE/g dry weight (Benavides-Guerrero et al., 2020; Navarro et al., 2020).

The extraction of these compounds from the byproducts of cider processing (called magaya) was optimized, and the concentration of polyphenols in two sample types was determined. The results showed that the total polyphenols in both types of magayas ranged from 1.36 to 6.39 mg/g, with flavonols comprising 50%–92% and hyperine predominating. The concentration of hyperine was between 0.14 and 0.42 mg/g for industrial magayas. In addition, polyphenols were extracted from the byproducts of broccoli processing (stems and leaves), obtaining a concentration of phenolic compounds ranging from $4,503.80 \pm 101.33$ to $17,630.47 \pm 539.34$ µg GAE/g extract. In the winemaking (seeds, skins, and stems) of the Zalema grape variety, the seeds had a higher content of total polyphenols (9829.86 mg GAE/100 g) than the stems (3745.77 mg GAE/100 g) and the skins (2797.67 mg GAE/100g). For the processing of pisco (residual grape seeds of eight varieties), the results indicated that between 402 and 84 mg EAG/g of seed could be obtained (Borja, 2023; Surco et al., 2020).

3.2 Carotenoids

Studies have extracted carotenoids from passion fruit shells. One study obtained 1037.99 ± 48.70 µg of β -carotene/100g, while another study reported 3.86 mg of β -carotene/100 g. Amazonian fruits such as chontaduro (*Actris gasipaes*) and ombu (*Phytolacca dioica* L.) have been studied, achieving 151.50 mg/100 g and 2,751 µg/100 g, respectively. This represents potential for obtaining this bioactive compound (Molina et al., 2019; Tarazona et al., 2020).

Moreover, this compound was extracted from dried pumpkin pulp waste, obtaining 613.9 ± 19.50 µg β -carotene equivalent/g. It was also extracted from agrifood byproducts, such as red grape pomace, edible flower petals, and plum tomato peels. Notably, five carotenoids were identified: lutein, β -cryptoxanthin, α -carotene, β -carotene, and 13-cis- β -carotene. Among these, β -carotene is the

predominant carotenoid (Jofre et al., 2020; Portillo et al., 2021).

Lycopene is an acyclic carotene extracted from the peels of red fruits and vegetables, such as tomatoes, red peppers, and watermelons. Flores et al. (2021) extracted lycopene from the byproducts of tomato product processing (seeds, peels, and fiber). The concentrations ranged from 119.2 ± 2.5 to 147.1 ± 6.8 mg lycopene/kg of dry sample with 7% moisture.

4. Extraction methods

4.1 Conventional

The conventional extraction method has been widely used for extracting compounds of interest from agroindustrial waste for many years. Although new technologies have appeared, it remains a viable option due to its efficiency, often outperforming most unconventional methods.

Using this method, various compounds have been extracted from fruit peels, including anthocyanins from red dragon fruit peels, phenolic compounds from pomegranate peel extract, and pectin from orange, passion fruit, and cucumber peels (Cabrera et al., 2019; Coca, 2021; 2020; Torres-Mendoza et al., 2023).

In addition, the conventional method has been used to extract bioactive compounds from other agroindustrial byproducts, such as phenolic compounds from the processing of pulp and baru nut (*Dipteryx alata*) as well as from strawberry processing and carotenoids from coffee processing byproducts Red and Yellow Caturra (Viana et al., 2023; Villamil-Galindo et al., 2021).

4.2 Unconventional extraction methods

Enzymatic method: The benefits of this method include its high specificity, catalytic activity, and the fact that it does not require elevated temperatures or pressures. It is environmentally friendly as it does not produce volatile components and is used to extract phenolic compounds, oils, pectin, and more. However, its primary disadvantage is that the enzymes require extended periods to act on the extracted compounds (Alavarsa, 2022). Pectin has been extracted from mango peels and sugar beet byproducts using this method (Pacheco, 2019).

Solid-State Fermentation (SSF): Solid-state fermentation (SSF) is an alternative biotechnological approach for extracting bioactive compounds from agroindustrial waste. It combines conventional and unconventional methods and is considered a clean technology. Its efficiency depends on the fungal species, the substrate used, and the environmental

conditions. In some research, agroindustrial byproducts have been used as substrates (called agrosubstrates) to generate conditions for fungal growth. These include red chickpea shells, rice and wheat bran, pineapple and apple residues, and sugarcane bagasse (Vargas-Sánchez et al., 2022).

Paz (2022) extracted phenolic compounds from the heart and peels of pineapple with the help of *Aspergillus niger* GH. Some phenolic compounds identified in this study were caffeoyl hexoxide and p-coumaroyl hexoxide. In another study, onion peas, potato, bean, and pea pods were used as substrates to produce carotenoids with *R. mucilaginosa*. The extracted carotenoids included β -carotene, phytoene, torulene, and torularhodin (Sharma & Ghoshal, 2020).

Ultrasound Assisted: Ultrasound-assisted extraction is an unconventional technique utilized in the agrifood industry because of its rapidity and effectiveness in inhibiting or reducing heat-resistant microorganisms. The foundation of this method relies on the use of high-frequency, low-intensity sound waves that do not cause harm or invasiveness to food (Corilla, 2019). Using this method, bioactive compounds have been extracted from agroindustrial byproducts, including polyphenols from coffee processing byproducts, yellow dragon fruit husk, santy shell, and avocado seed and peel. The extraction of anthocyanins from jaboticaba (*Plinia cauliflora*) shells (a native berry from Brazil) was also optimized. The research indicates that the extraction time variable is more substantial than the solvent concentration and temperature. Compared to ultrasound extraction, heat-assisted extraction is the most effective method for extracting anthocyanins from berry peel, with cyanidin-3-O-glycoside being the main anthocyanin (Albuquerque et al., 2020; Corilla, 2019; Monzón et al., 2021; Rojas et al., 2019; Torres-Valenzuela et al., 2020).

Microwave Assisted: Microwave-assisted extraction is an eco-friendly technique in the agrifood industry, utilizing an electromagnetic field to increase cell breakage and reduce bioactive compound degradation (Figueroa et al., 2021). The method for extracting anthocyanins and phenolic compounds from elderberry shells supports the theory that lower irradiation power and higher solvent concentration improve performance. Extraction time is excessive as an equilibrium between solute and solvent can cause degradation. Other phenolic compounds were extracted from avocado shells and pectin from apple peel under different conditions, including temperature, potential of hydrogen (pH), and extraction

time. High-performance liquid chromatography (HPLC) was used to quantify and identify compounds (Figueroa et al., 2021).

Supercritical Fluids and Pressurized Liquids (PLE): Green extraction techniques involve using solvents with varying properties, such as viscosity and diffusivity, to improve the efficiency of the extraction process. Supercritical fluids, when subjected to temperature and pressure conditions above the critical point, demonstrate different properties, making them easier to introduce to the sample. Carbon dioxide is commonly used for its safety, low cost, and low critical points. In addition, PLE extraction is time-efficient, safe, and fast, using high pressure to keep the solvent liquid until it boils. It also protects the sample from light and oxygen (Borja, 2023). Fatty acids have been extracted from oil obtained from tamarind and mango seeds with supercritical fluids. Similarly, polyphenols, carotenoids, and other bioactive ingredients have been extracted from olive oil byproducts (Cerón-Martínez et al., 2021; Difonzo et al., 2021; Idris et al., 2022). However, Rubiano-Charry et al. (2019) showed that extracting bioactive compounds (polyphenols and vitamin C) from mango peels using supercritical fluids was not feasible on a large scale due to their low yield and long processing times. The study suggests that pressurized liquid extraction is unsuitable for industrial use but recommended for laboratory or small-scale applications. This method has been used to extract phenolic compounds from pomace, resulting in extracts with increased content and selective identification. It is used to extract glycoalkaloids from native potato peels (Table 1) (Cea, 2020; Eraso-Grisales et al., 2019).

Deep Eutectic Solvent (DES) extraction: Deep eutectic solvent (DES) extraction involves mixtures with lower melting points and transition temperatures, making them an excellent alternative to organic solvents. They comprise a hydrogen bond acceptor and donor and are considered green or smart solvents. Notably, DES with natural constituents (natural deep eutectic solvents [NADES]) are also used to extract bioactive compounds from agroindustrial waste, achieving high efficiency, purity, and yield. (Saini et al., 2022). Phenolic compounds have been extracted from ripe mango peels using a lactic acid/sodium acetate/water mixture in a ratio of 3:1:4 through microwave-assisted extraction and from brewer's bagasse using choline chloride (ChCl) and microwave-assisted glycerol (Figure 1) (López et al., 2021; Pal & Jadeja, 2020). The development of conventional and non-conventional extraction methods over time is shown in Figure 2.

Table 1

Summary of research on various extraction methods for bioactive compounds from agroindustrial byproducts

Extraction Method	Type of Waste	Compounds of Interest	Conditions	Results	Application/Potential use	References
Conventional	Orange peels	Pectin	Acid hydrolysis - pH of 3.2. Extraction time: 75 min, T: 80 °C	High degree of esterification. Yield: 20.4%	Food additive	Cabrera et al. (2019)
	Pomegranate peel	Phenolic compounds	Maceration in ethanol:water (38:62 v/v). Extraction time: 3.2 h	124.3 ± 2.9 mg *GAE/g	Unpasteurized fruit juice	Coca (2021)
	Cucumber peel	Pectin	Acid hydrolysis - pH of 1.5. Extraction time: 60 min, T: 85 °C	% Yield: 0.65%	Food additive for compotes	Torres-Mendoza et al. (2023)
	Strawberry processing byproducts	Phenolic compounds	1:10 solid:liquid ratio sequential extractions with 80% methanol + 0.5% formic acid. Centrifuged at 12,000 g for 20 min	30.04 g *GAE/Kg	Potential application in the nutraceutical industry	Villamil-Galindo et al. (2021)
	Byproducts of baru processing (pulp and walnut)	Phenolic compounds, flavonoids, and tannins	Ethanol: water (70:30, v/v)	529.50 ± 12.44 mg *GAE/100 g total phenolics, 29.47 ± 1.76 mg QE/100 g flavonoids	Cookie ingredient	Viana et al. (2023)
	Liquid effluents from vanilla and wine processing	Phenolic compounds	Ammonium salts, menthol, and fatty acids. 2-methyltetrahydrofuran, cyclopentyl methyl ether, and limonene	Extraction values close to 100%. Extraction potential of natural antioxidants from aqueous effluents	Potential application in the food industry	Cañadas Soler (2022)
	Yellow dragon fruit peel	Total polyphenols	Frequency: 35 kHz, rated power: 222 W. Amplity: 60%. Sonication time: 22 min	973.10 mg/L Gallic Acid	Potential application in the pharmaceutical, cosmetic, or nutraceutical industries	Torres-Valenzuela et al. (2020)
Ultrasound Assisted	Sanky peel	Phenolic compounds	Frequency: 20 Hz. Power: 130 W. Sonication time: 40 min. T: 25 °C with 50% ethanol	43.9 mg *GAE/g	Potential application in the food industry	Rojas et al. (2019)
	Coffee processing byproducts (pulp residues)	Total polyphenols	Ethanol 55%. Frequency: 95%. Extraction time: 9 min	41.1646 mg *GAE/g	Potential application in the food and pharmaceutical industries	Corilla (2019)
	Avocado seeds and peels	Total polyphenols	Power: 100 W. Frequency: 42 Hz. T: 49.5 °C, 42% ethanol for 65.5 min (seeds); T: 51.1 °C, 49.5% ethanol for 62 min (shells)	145.17 mg *GAE/g (seeds), 124.05 mg *GAE/g (shells)	Potential application in the food industry	Monzón et al. (2021)
	Jaboticaba husk	Anthocya-nins	Power: 500 W. Frequency: 20 Hz. 34.47% ethanol (v/v)	31 ± 2 mg/g extract	Natural colorant in a bakery product	Albuquerque et al. (2020)
Microwave Assisted	Avocado peel	Total polyphenols	T: 130 °C per 39 min, ethanol 36%, solvent:sample ratio: 44 mL/g	72.04 mg *GAE/g	Natural food preservative, functional food ingredient, or nutraceutical	Figuerola et al. (2021)

Enzymatic Extraction	Sugar beet byproducts	Pectin	Commercial <i>Trichoderma reesei</i> pH of 4.5 (1:20 w/v) cellulase, continuous stirring: 125 rpm T: 61 °C for 102 min. Resting at room temperature for 24 h	High galacturonic acid content: 72%, Yield: 13%	Food additive	Pacheco (2019)
	Tamarind seed oil	Fatty acids	CO ₂ flow: 4 mL/min. Extraction time: 45 min, T: 80 °C, P: 7000 psi	Yield: 0.34%	Raw material for emulsifiers and coatings	Idris et al. (2022)
	Mango seed oil	Essential fatty acids	30 g CO ₂ /min through fixed bed for 150 min, P: 30 MPa and T: 63 °C	37 g/kg linoleic acid, 4 g/kg α-linolenic acid, 155 g/kg oleic acid	Potential application in the cosmetic, pharmaceutical, and food industries	Cerón-Martínez et al. (2021)
Supercritical Fluids	Olive oil byproducts	Carotenoids, total polyphenols, and others	Q: 1L CO ₂ /min, T: 40°–60 °C, P: 250–350 bar, 3 min static cycle and 10–15 min dynamic cycle	259.9–424.5 mg/kg α-tocopherol, 3450–4316 mg/100 g squalene, 23.9–198.9 mg/100 g β-Sitosterol	Potential application in the food and pharmaceutical industries	Difonzo et al. (2021)
	Mango peel	Total polyphenols	P: 300 bar, T: 40 °C, CO ₂ flow: 13 mL/min. Static extraction: 15 min. Dynamic extraction: 75 min	Yield: 56.67%	Potential application in the food industry	Rubiano-Charry et al. (2019)
Pressurized Liquids	Native potato peel	Glycoalkaloids	P: 80 bar, T: 80 °C for 40 min	0.784 mg/g α-solaninel, 637 mg/g α-chaconine, 2.420 mg **GAT /g	Potential application in the pharmaceutical industry	Eraso-Grisales et al. (2019)
	Byproduct of olive oil processing (pomace)	Phenolic compounds	P: 1500 psi per 20 min 52.3% ethanol, T: 136.5 °C	1659 mg/kg	Potential application in the nutraceutical industry	Cea (2020)
Solid-state fermentation	Onion peels, potato peas, bean peas, and pea pods	Carotenoids (β-carotene)	<i>Rhodotorula mucilaginosa</i> , pH of 6.1, T: 25.8 °C and stirring 119.6 rpm for 84 hrs	17.35 µg/g β-carotene, 819.23 µg/g total carotenoids	Natural colorant	Sharma & Ghoshal (2020)
	Heart and pineapple peels	Phenolic compounds	<i>Aspergillus niger</i> GH1 26% ethanol Solid/liquid ratio: 82 mg/mL, cycles:2	12 mg *GAE/g	Potential application in the food, cosmetic, and pharmaceutical industries	Paz (2022)
Deep Eutectic Solvents	Ripe mango peels	Phenolic compounds	DES: Lactic acid/sodium acetate/water (3:1:4) P:436.45 W, t:19.66 min, liquid-to-solid ratio: 59.82 mL/g	56.17 mg *GAE/g	Potential application in the food and pharmaceutical industries	Pal & Jadeja (2020)
	Beer Bagasse	Total polyphenols	DES: ChCl: glycerol, T: 100 °C for 13.30 min, 37.46% (v/v) water at DES	2.89 mg *GAE/g	Application in a biorefinery (biofuels or chemicals)	López-Linares et al. (2021)

*mg GAE/g: milligram of gallic acid equivalent per gram.

**mg GAT/g: milligram of total glycoalkaloids per gram.

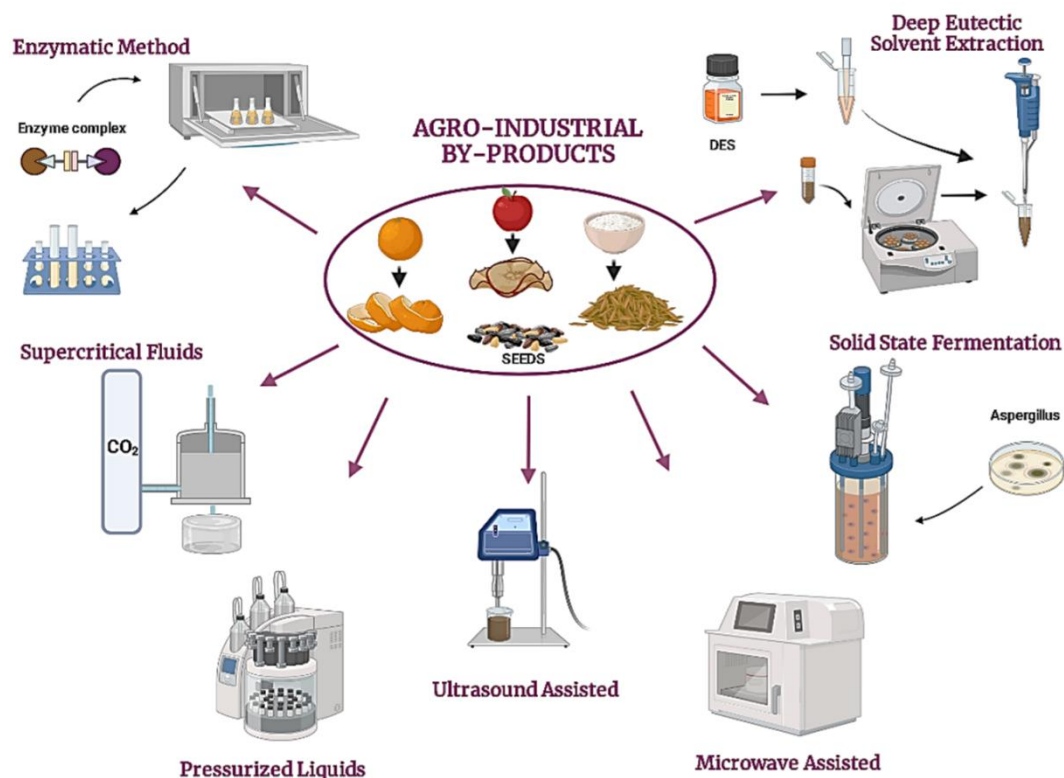


Figure 1. Unconventional extraction methods.

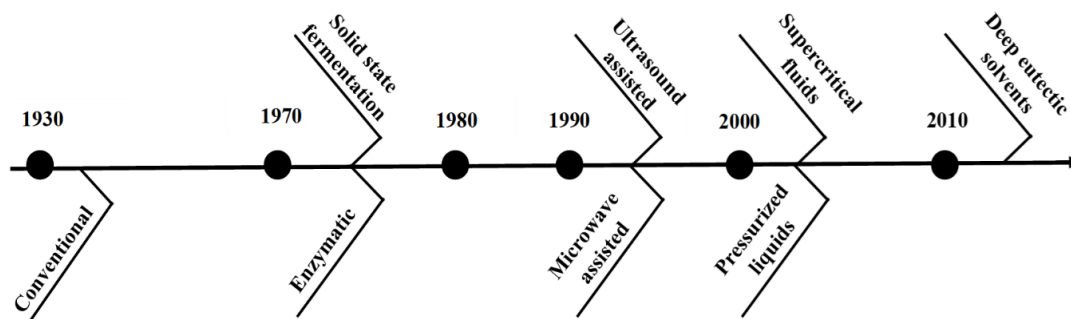


Figure 2. Schema of extraction methods timeline.

5. Microencapsulation methods

Spray drying or atomization drying: Microencapsulation by spray drying is a widely used method in the agrifood industry for protecting bioactive compounds from environmental factors, such as temperature, pH, light, relative humidity, and oxygen, thereby extending their shelf life. This method enhances stability under processing, storage, and gastrointestinal tract conditions. It is cost-effective and widely available. The process involves atomizing a liquid suspension coated with a polymer layer in a high-temperature chamber, causing the solvent to evaporate quickly and trapping the bioactive compound inside the encapsulating material (Figure 3) (Navarro et al., 2020). This method has been used to micro-

encapsulate phenolic compounds from coffee and cocoa processing residues, anthocyanins from senescence blackberries and purple cabbage leaves, and lycopene from tomato residues (Corilla, 2019; Espinosa et al., 2019).

Freeze-drying or lyophilization: Freeze-dried microencapsulation is used for dehydrating and microencapsulating foods containing heat-sensitive compounds, such as anthocyanins. It involves freezing, sublimation, desorption, and storage stages. The encapsulating material is mixed with the core compound, homogenized, and stored for freeze-drying. An extract of leaves soursop was microencapsulated with phenolic compounds, maltodextrin, and gum Arabic (Jordan, 2019). A study on blueberry processing byproducts found

that the productivity of microcapsules in anthocyanin extract is influenced by the concentration and type of encapsulating agent (Castagnini et al., 2019).

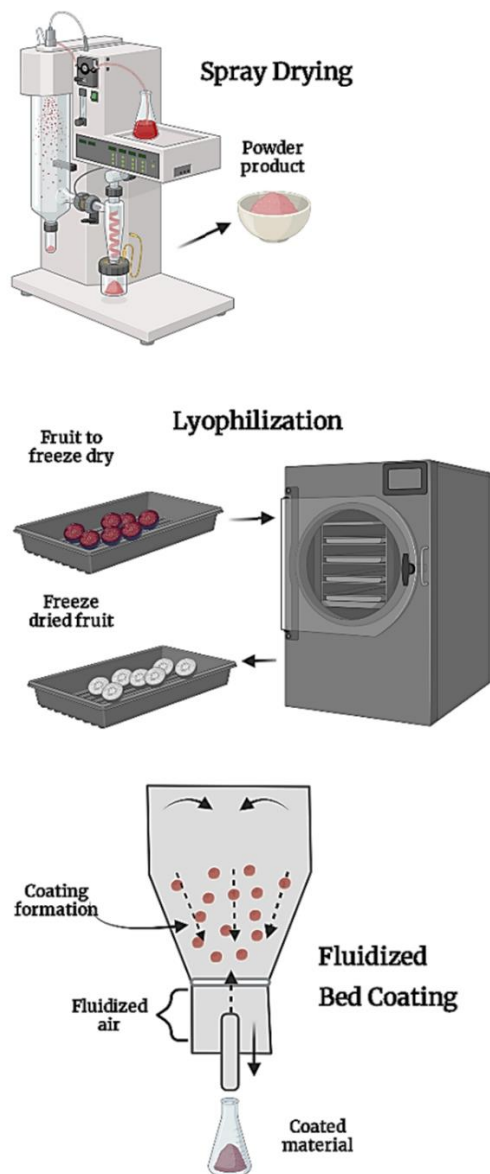


Figure 3. Microencapsulation methods.

Fluidized bed coating: Microencapsulation involves applying a coating to powder microparticles, either continuously or discontinuously. Factors such as the solid's circulation speed, nozzle atomization pressure, coating feeding speed, and temperature affect agglomeration, film formation, and coating efficiency. This technique is applicable to meltable coatings, such as hydrogenated vegetable oils, fatty acids, waxes, and emulsifiers, or soluble coatings, such as maltodextrins, gums, and starches. In

meltable coatings, cold air is used to harden the carrier, while in soluble coatings, hot air evaporates the solvent, releasing its content when water is added. Phenolic compounds from pomace and alpechin were extracted and microencapsulated using spray drying and a fluidized bed. The coated microparticles showed reduced solubility as particle size increased, indicating that the fluidized bed provides site-specific release properties. This application also allowed the coating of surface polyphenols, increasing the encapsulation efficiency of some compounds (Cea, 2020). The initial polyphenol value in Hass avocado peels and seed extract decreased with spray drying microencapsulation but increased after coating with modified starch, even in greater quantities than in the extract (Table 2) (Asenjo, 2018).

Nanoencapsulation: Nanotechnology involves trapping active ingredients in a roof wall, making them resistant to external factors, such as light, temperature, and pH. This enhances control over their release. Nanoencapsulation allows for the inclusion of hydrophobic compounds in beverages and cosmetics due to their enlarged surface area. Green nanochemistry is used to synthesize metal nanoparticles, avoiding toxic chemical reagents (Figure 4). Nanoparticles from agroindustrial byproducts are more biocompatible than synthetic polymers (Meral et al., 2022). Studies have found that using ionic gelation, nanoencapsulated polyphenols from artichoke and beet leaves maintained their antioxidant activity (Espinoza & Ricanqui, 2022). Bioactive peptides from jackfruit seed have also been nanoencapsulated in solid lipid nanoparticles to be used as food additives and nutraceuticals (Table 3). The development of encapsulation methods over time is shown in Figure 5.

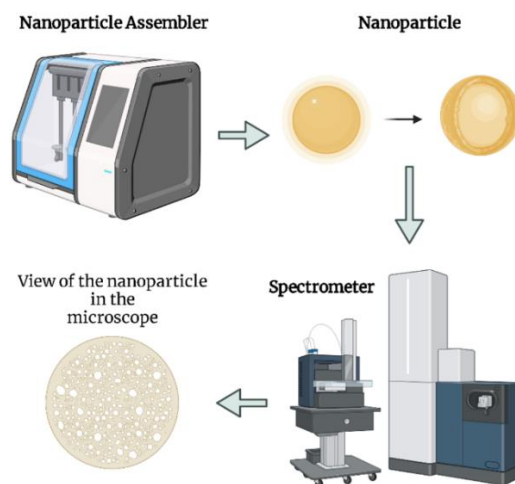


Figure 4. Nanoencapsulation method schema.

Table 2

Summary of research on different types of microencapsulation of bioactive compounds from agroindustrial byproducts

Microencapsulation Method	Type of Waste	Compounds of Interest	Conditions	Results	Potential use	References
Spray Drying	Coffee Processing Waste	Total polyphenols	Maltodextrin 7% Inlet air T°: 120 °C	30.32 ± 0.19 mg GAE/g	In vitro application in different topics on health	Corilla (2019)
	Purple cabbage leaves	Anthocyanins	Maltodextrin-anthocyanin concentrate ratio (2:1) Inlet temperature: 150 °C, Outlet temperature: 90 °C	Yield: 58.9%	Ingredients in fermented milk drinks	Espinosa et al. (2019)
Freeze-drying	Soursop Leaf Extract	Phenolic compounds	Maltodextrin 10% Chamber T°: 140 °C. Feeding luxury: 7 mL/min	Encapsulation efficiency: 77.68 ± 0.62%	Application for therapeutic purposes	Jordan (2019)
	Byproduct of cran-berry juice processing (solid re-mains of the skin)	Anthocyanins	Encapsulating matrix : Whey Protein–Maltodextrin (70:30)	Encapsulation efficiency: 87%–99%	Potential application in the food and pharmaceutical industries	Castagnini et al. (2019)
Fluidized Bed Coating	Byproduct of olive oil processing (pomace and alpechín)	Total polyphenols	Coating with inulin (10% w/v) and so-dium alginate (2% w/v) per fluidized bed for 2 h at 50 °C	1.4 mg EAG/g (with sodium alginate), 1.5 mg EAG/g (with inulin)	Potential application in the food and pharmaceutical industries	Cea (2020)
	Hass avocado peels and seeds	Total polyphenols	Coating with modified corn starch at 35 °C tempera-ture for 20 min.	2117.9 mg EAG/g peels, 3867.3 mgEAG/g seed	Potential application in the pharma-ceutical industry	Asenjo (2018)

Table 3

Summary of research on nanoencapsulation of bioactive compounds from agroindustrial byproducts

Type of Waste	Compounds of Interest	Conditions	Results	Potential use	References
Artichoke waste (bracts)	Total polyphenols	Ionic gelation: Chitosan concentration: 0.28% sodium triphosphate concentration: 0.29% ratio to crosslinking agent: 1/5 pH: 4.9 and sonication time: 4.79 min	Optimal encapsulation efficiency: 69.9%	Applications for food and therapeutic purposes	Pacheco (2019)
Jackfruit seeds	Bioactive Peptides	With solid lipid nanoparticles (NLS), 4.5% cocoa butter 0.5% beeswax	Encapsulation efficiency: 85.96 ± 1.13%	Application in the development of functional foods	Cruz (2023)
Beet greens	Total polyphenols	Ionic gelation: Chitosan: 0.136 % Triphosphate: 0.2% Q/TTP ratio: 4.99%, pH: 3.0 and sonication time: 0.0031 min	Optimal encapsulation efficiency: 62.71 %	Potential application in the food and pharmaceutical industries	Espinosa & Ricanqui (2022)

6. Applications and possible uses of bioactive compounds from agroindustrial waste

Studies have explored the use of bioactive compounds from agroindustrial byproducts in food for various purposes, including shelf-life extension, improved organoleptic, and functional or nutraceutical properties (Table 4). Ordóñez-Santos et al. (2022) used ultrasound-assisted extraction of total carotenoids from papaya peels to extract total carotenoids from Frankfurt sausages. The extract was used as a natural coloring additive during storage, and after 30 days, the sausage retained its initial color, smell, and taste. Rojas et al. (2022) extracted, encapsulated, and added carotenoids obtained from the byproducts of the processing of Arabica coffee of the Red and Yellow Caturra varieties to a dairy product

(yogurt). Three yogurt samples were emulsified with microencapsulated extract, two of which were stirred and stored at 4 °C. The extract improved the product's viscosity and color index, indicating its potential for application and demonstrating its feasibility. Rodríguez (2023) extracted phenolic compounds from pomegranate peel and added them to the preparation of chicken burgers in two concentrations (0.1% and 1.0%). The study analyzed burgers stored in refrigerated (4 °C) and frozen (−18 °C) conditions, measuring their pH, color, shear strength, TBARS, and protein levels at 0, 4, 8, 12, and 16 days. The results showed that ultrasound-assisted extraction of polyphenol extract inhibited lipid and protein oxidation and stabilized the characteristic color of the product.

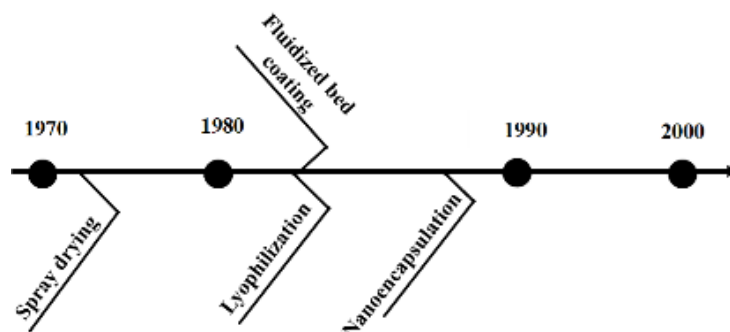


Figure 5. Schema of encapsulation methods timeline.

Table 4

Some applications in the food industry

Type of Residue/Compound of Interest	Application	Physicochemical Characterization	Sensory Characterization	References
Papaya Peel / Total Carotenoids	Preparation of Frankfurt sausages	Residual nitrite decreased from 41.35 to 21.66 mg/kg. Peroxide index increased from 23.50 to 29.78 meq O_2 /Kg. P-anisidine value increased from 25.18 to 28.57 μ mol/ μ g. Lipid oxidation increased from 0.41 to 0.51 mg MDA/kg	This substance prolongs shelf life by delaying lipid oxidation without causing unpleasant aromas or flavors, preserves color during storage, and may partially replace nitrates.	Ordóñez-Santos et al. (2022)
Arabica Coffee Byproducts/ Total Carotenoids	Making fortified yogurt	Viscosity increased, L Reference: 85.8 and Sample: 77.67, a Reference: -0.6 and Sample: 4.18, b Reference: 13.33 and Sample: 15.18	More reddish and yellowish and slightly bitter than the reference	Rojas et al. (2022)
Pomegranate Peel / Phenolic Compounds	Preparation of chicken burgers	Increased cooking performance. Lipid oxidation: Frozen burger: 0.22 mg MDA/kg, chilled: 1.50 mg **MDA/kg, Protein oxidation: Frozen burger: 1.94 nmol carbonyl/mg protein, chilled: 3.56 nmol carbonyl/mg protein	Improves organoleptic properties and characteristics considerably in frozen storage	Rodríguez (2023)

** mg MDA/kg: milligram of malondialdehyde per kilogram.

The food industry is incorporating bioactive compounds from agroindustrial waste into packaging due to their commercial and technological potential. Biopolymers, including polysaccharides, proteins, and phenolic compounds, are extracted from plant sources such as potatoes and cereals. Starch, obtained from potatoes, rice, corn, and wheat, offers antioxidant activity, is low-cost, and is the richest biomaterial. Carotenoids are also added to biofilms used in food packaging production. The optimal use of β -carotene is achieved by encapsulating it before adding it to active biodegradable packaging films. A packaging for sunflower oil made of lactic acid films with lycopene, β -carotene, and bixin (a natural dye from annatto seeds) proved to be an effective barrier against oxygen and light, with enhanced antioxidant activity for the oil. Synthetic stabilizers have been replaced by lycopene and other active ingredients obtained from byproducts of tomato and grape processing. Chlorophyll is used as a colorimetric temperature indicator (50–70 °C) in chitosan films, while

anthocyanin can detect temperature variations in smart packaging as it changes color with temperature (Nemes et al., 2020).

7. Applications in other industries

The cosmetic industry is interested in agroindustrial byproducts such as lycopene, which possess bioactive compounds with photoprotective properties for the skin. Lycopene, obtained from tomato processing residue, maintains cell proliferation and protects the skin from ultraviolet (UV) damage by downregulating epidermal ornithine decarboxylase activity. As a key ingredient in microemulsions, lycopene stimulates antioxidant activity in tissues. Nanoparticle formulations have been developed to stabilize lycopene in cell culture mediums, thereby enhancing cell uptake. Oleuropein, a phenol from olive processing residue, scavenges free radicals, protects against UVB rays, and inhibits skin elasticity and thickness loss.

Quercetin, a flavonol from apple processing by-products, is an anti-inflammatory agent. However,

its dermal absorption capacity is limited. Therefore, incorporating it into glycosomal and liposomal nanoformulations is necessary to enhance its ability to eliminate free radicals and protect skin tissue from damage caused by UVB irradiation. Ellagic acid (obtained from the byproducts of pomegranate processing) is a phenolic acid that blocks the infiltration of inflammatory macrophages into integuments and decreases the production of proinflammatory cytokines (Simitzis, 2018).

Aquaculture is exploring the use of bioactive compounds from agroindustrial waste as alternatives to antibiotics and prebiotics due to their antimicrobial and antioxidant properties as well as immune system modulation benefits. This approach aims to enhance production chains, reduce pollution, and enhance the well-being of organisms. For example, phenolic compounds obtained from the processing of wine (grape seeds) and olive oil have potential as food additives due to their immunostimulant properties. Terpenes obtained from citrus peels are an alternative source of essential oil to antibiotics, allowing for a 48.33% increase in fish survival under conditions of high disease exposure. Research indicates that prebiotics can be obtained from agroindustrial residues (such as wheat), fruits (such as bananas, mangoes, and grapes), and citrus fruits (such as oranges and lemons) (Leyva-López et al., 2020).

The use of agroindustrial byproducts in animal feed and nutrition has integrated these byproducts into livestock and agricultural systems. Fruit and vegetable processing waste can be used as animal feed concentrates, reducing grain consumption in plant food. Bioactive compounds also benefit the environment by reducing methane and nitrogen emissions, while animal-based foods increase their nutraceutical value (Correddu et al., 2020).

Grape seed extract enriched with flavan-3-ol has been found to effectively inhibit the growth of lactic acid bacteria in the bioethanol industry, outperforming yeasts. However, due to limited research, it is recommended to minimize the use of hazardous chemicals in combating bacterial contamination (Shirahigue & Ceccato, 2020).

8. Future perspectives

The bioavailability of bioactive compounds from agroindustrial byproducts directly impacts health. Technological advances enhance absorption in the gastrointestinal tract and gut microbiota, leading to beneficial health effects. Clinical trials examining the relationship between noncommunicable diseases, such as obesity and metabolic syndrome, under-

score the importance of revaluing agroindustrial waste and evaluating its nutraceutical, functional, physiochemical, microbiological, and sensorial properties (Jeria et al., 2022; Reguengo et al., 2022). A recent study enabled the production of functional cereal bars from wheat glucose syrup obtained through the SSF of wheat bran. These rice flake bars had better nutritional value than commercial bars, demonstrating that SSF remains an attractive option (Figueiredo et al., 2024).

Shelf-life studies of extracts from fruit peels, such as pineapple, obtained using conventional methods have provided information on their shelf life, ranging from 40 to 60 days depending on environmental conditions. It is important to complement these studies with microbiological analyses to assess the factors that may influence the stability of the extracts. Furthermore, applying green technologies in the extraction process is essential as they offer the potential for higher yields and sustainability advantages than conventional technology. New generations of colorants are being developed from sediments of beverage processing, such as wine, for applications in various industries, including food and cosmetics. These colorants possess various characteristics, including color intensity, flavor, shelf life, and bioavailability, which pose challenges for future developments (Armas Proaño & Ashqui Silva, 2024).

A brief bibliometric analysis using the search criteria TITLE-ABS-KEY: "Agroindustrial," "Waste," and "Bioactive-compounds" highlighted published articles and helped to detect gaps in knowledge or areas requiring further exploration on this topic. Keyword co-occurrence mapping was performed using a VOSviewer. Figure 6 (a) shows the network visualization chart in which five clusters are identified. The larger red cluster represents the focus of published articles on "bioactive compounds" related to "fermentation" (Ydav et al., 2024); the yellow cluster is related to the "food processing" industry that generates the most waste. The green cluster is related to the antioxidant activity of the byproducts of the agrifood sector. The blue cluster is related to antioxidants and their extraction methods. Finally, the purple cluster is linked to agroindustrial wastes, which are being studied for their benefits in sustainability and the circular economy (Chiaraluce et al., 2021). In addition, Figure 6 (b) shows the topics arranged chronologically, with the most recent ones in yellow nodes and the old ones in purple. Older keywords encompass both solid and liquid waste, while more recent topics include waste recovery, Bioconversion, and biological activity.

Contribution by authors

J. Floridas-Galán: Conceptualization, Methodology, Software, and Writing—original draft, review, and editing. **P. Paredes-Concepción:** Formal analysis, review, editing, and supervision. All authors have read and agreed to the published version of the manuscript.

Conflicts of interests

The authors declare no conflicts of interest.

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References

- Aguilar-Méndez, M. A., Campos-Arias, M. P., Quiroz-Reyes, C. N., Ronquillo-de Jesús, E., & Cruz-Hernández, M. A. (2020). Fruit peels as sources of bioactive compounds with antioxidant and antimicrobial properties. *Revista de la Facultad de Ciencias Agrarias UNcuyo*, 52(1), Article 1.
- Alavarsa, D. (2022). *Desarrollo y optimización de la extracción enzimática y análisis de compuestos con actividad biológica en açai (Euterpe oleracea Mart.)*. Trabajo de grado. Universidad de Cádiz. España.
- Albuquerque, B. R., Pinela, J., Barros, L., Oliveira, M. B. P. P., & Ferreira, I. C. F. R. (2020). Anthocyanin-rich extract of jabuticaba epicarp as a natural colorant: Optimization of heat- and ultrasound-assisted extractions and application in a bakery product. *Food Chemistry*, 316, 126364. <https://doi.org/10.1016/j.foodchem.2020.126364>
- Angulo Arias, L. V., De Souza Silva, V., Augustus de Oliveira, R., & Matta Fakhouri, F. (2018). Caracterización de subproductos agroindustriales: Naranja y maracuyá. *Ingeniería y Región*, 20, 59-66. <https://doi.org/10.25054/22161325.1916>
- Armas Proaño, W. U., & Ashqui Silva, A. R. (2024). *Obtención de un colorante a partir de los sedimentos producidos en la elaboración de vino de mortiño*. Bachelor Thesis, Ecuador: Latacunga: Universidad Técnica de Cotopaxi (UTC).
- Aseño, A. C. (2018). Desarrollo de extractos secos con propiedades antioxidantes y antibacterianas de cáscara y semilla de palta Hass (*Persea americana*) recubiertos con almidón modificado. Tesis de Magister, Universidad de Chile.
- Astudillo, Á., Rubilar, O., Briceño, G., Diez, M. C., & Schalchli, H. (2023). Advances in Agroindustrial Waste as a Substrate for Obtaining Eco-Friendly Microbial Products. *Sustainability*, 15(4), 3467. <https://doi.org/10.3390/su15043467>
- Bamigbade, G. B., Subhash, A. J., Tarique, M., Al-Ramadi, B., Abu-Jdayil, B., Kamal-Eldin, A., Nyström, L., & Ayyash, M. (2024). Date pomace polysaccharide: Ultrasonic-assisted deep eutectic solvent extraction, physicochemical properties, biological activities, gut microbiota modulation, and rheological properties. *Chemical and Biological Technologies in Agriculture*, 11(1). <https://doi.org/10.1186/s40538-024-00601-0>
- Benavides-Guerrero, R., Revelo-Cuarán, Y. A., Arango-Bedoya, O., & Osorio-Mora, O. (2020). Extracción asistida con ultrasonido de compuestos fenólicos de dos variedades de papas (*Solanum phureja*) nativas andinas y evaluación de su actividad antioxidante. *Información Tecnológica*, 31(5), 43-50. <https://doi.org/10.4067/S0718-07642020000500043>
- Borja Martínez, M. (2023). Aprovechamiento de los subproductos del brócoli como fuente de compuestos bioactivos. *Proyecto de Investigación*. Doctoral Thesis. Universidad de Murcia. España.
- Bustillos-Sánchez, D. L., & Cochero Murillo, B. L. (2022). Obtención de una bebida alcohólica destilada (vodka) mediante hidrólisis ácida a partir de almidón de oca (*Oxalis tuberosa*). Thesis, Universidad Politécnica Estatal del Carchi. Ecuador.
- Cabrera, T. A. P., Armas, S. E. C., Almeida, C., & Carrillo, I. (2019). Diseño de una planta piloto de extracción de pectina como gelificante a partir de residuos de la naranja (*Citrus sinensis*). *FIGEMPA. Investigación y Desarrollo*, 8(2), Article 2. <https://doi.org/10.29166/revfig.v1i2.1274>
- Cañadas Soler, R. (2022). Valorización de residuos agroindustriales mediante nuevos procesos sostenibles para la obtención de antioxidantes naturales [PhD, E.T.S.I. Industriales (UPM)]. <https://oa.upm.es/71730/>
- Capillo Herrera, N., & Navarro Valdez, K. K. (2019). Efecto de la adición del extracto hidroetanólico de semilla de ungurahui (*Oenocarpus bataua* Mart.) en forma libre y microencapsulado sobre la calidad de la galleta. Tesis. Universidad Nacional Mayor de San Marcos.
- Casas-Rodríguez, A. D., Ascacio-Valdes, J. A., Davila-Medina, M. D., Londono-Hernandez, L., & Sepulveda-Torre, L. (2024). Use of pineapple (*Ananas comosus* L.) waste and its application in different biotechnological sectors. En *Waste Biorefineries*, (205-228).
- Castagnini, J. M., Rasia, M. C., Scattolaro, O. M., Capodoglio, D. L., Agout, M. F., Chacón, C., & Gerard, J. A. (2019). Microencapsulación de compuestos bioactivos. *Ciencia, Docencia y Tecnología Suplemento*, 9(9), Article 9.
- Cea, I. A. (2020). Extracción de compuestos fenólicos de subproductos provenientes de la industria del aceite de oliva y su estabilización mediante microencapsulación. Tesis. Universidad de Chile.
- Cerón-Martínez, L. J., Hurtado-Benavides, A. M., Ayala-Aponte, A., Serna-Cock, L., & Tirado, D. F. (2021). A Pilot-Scale supercritical carbon dioxide extraction to valorize Colombian mango seed kernel. *Molecules*, 26(8), Article 8. <https://doi.org/10.3390/molecules26082279>
- Cervantes-Sierra, R., Barragán-Condori, M., & Chaquilla-Quilca, G. (2019). Evaluación de antioxidantes en el té de hojas de camote morado (*Ipomoea batatas* L.). *Revista Tecnología en Marcha*, 32(4), 51-59. <https://doi.org/10.18845/tm.v32i4.4790>
- Chiaraluce, G., Bentivoglio, D., & Finco, A. (2021). Circular economy for a sustainable agri-food supply chain: A review for current trends and future pathways. *Sustainability*, 13(16), 9294. <https://doi.org/10.3390/su13169294>
- Coca, E. R. (2021). Microencapsulación de extracto de cáscara de granada por secado por atomización y su efecto en la estabilidad microbiológica de jugo de fruta no pasteurizado. Tesis. Universidad de Chile.
- Corredor, Y. A. V., & Pérez, L. I. P. (2018). Aprovechamiento de residuos agroindustriales en el mejoramiento de la calidad del ambiente. *Revista Facultad de Ciencias Básicas*, 1(1), Article 1. <https://doi.org/10.18359/rfcb.3108>
- Coriat, R. I. (2022). Optimización de la extracción simultánea de proteínas Y compuestos bioactivos presentes en la semilla de Camu-Camu (*Myrciaria dubia* (HBK) Mc. Vaughn). Tesis. Universidad Nacional Intercultural de la Amazonia. Perú.
- Corilla, D. D. (2019). Extracción, microencapsulación y cuantificación de compuestos fenólicos asistida por ultrasonido a partir de residuos de café (*Coffea arabica* L.). Tesis. Universidad Nacional del Centro del Perú.
- Cruz, M. E. (2023). Nanopartículas lipídicas sólidas para la encapsulación y estabilización de péptidos bioactivos de la semilla de yaca (*Artocarpus heterophyllus* lam). Tesis. Universidad de Chile.
- Jeria, N., Cornejo, S., Prado, G., Bustamante, A., García-Díaz, D. F., Jimenez, P., Valenzuela, R., Poblete-Aro, C., & Echeverría, F. (2023). Beneficial effects of bioactive compounds obtained from agro-industrial by-products on obesity and metabolic syndrome components. *Food Reviews International*, 39(7), 3753-3782. <https://doi.org/10.1080/87559129.2021.2013498>
- Correddu, F., Lunesu, M. F., Buffa, G., Atzori, A. S., Nudda, A., Battacane, G., & Pulina, G. (2020). Can agro-industrial by-products rich in polyphenols be advantageously used in the feeding and nutrition of dairy small ruminants? *Animals*, 10(1), Article 1. <https://doi.org/10.3390/ani10010131>

- Difonzo, G., Aresta, A., Cotugno, P., Ragni, R., Squeo, G., Summo, C., Massari, F., Pasqualone, A., Faccia, M., Zambonin, C., & Caponio, F. (2021). Supercritical CO₂ extraction of phytocompounds from olive pomace subjected to different drying methods. *Molecules*, 26(3). <https://doi.org/10.3390/molecules26030598>
- Eraso-Grasales, S., Mejía-España, D., & Hurtado-Benavides, A. (2019). Extracción de glicoalcaloides de papa nativa (*Solanum phureja*) variedad ratona morada con líquidos presurizados. *Revista Colombiana de Ciencias Químico-Farmacéuticas*, 48(1), 181-197. <https://doi.org/10.15446/rcciquifa.v48n1.80074>
- Espinosa Álvarez, C. S., López Contreras, J., Escobar Rodríguez, D., Jiménez Rondón, D., Bugueño Muñoz, W., & Cereza Mezquita, P. (2019). Application of microencapsulated anthocyanin extracted from purple cabbage in fermented milk drinks. *Acta Agronómica*, 68(2), 134-141. <https://doi.org/10.15446/acag.v68n2.79078>
- Espinoza, A. R., Ricanqui, E. S. (2022). Nanoencapsulación por gelificación iónica de polifenoles a partir de hojas de beterraga (*Beta vulgaris* L.) asistidas por ultrasonido. Tesis de grado, Universidad Nacional del Centro del Perú.
- Figueiredo, C. C. M., Granero, F. O., Silva, L. P., Nogueira, I. F. A., de Souza, J. F., Escaramboni, B., de Oliva Neto, P., & da Silva, R. M. G. (2024). Solid-state fermentation using wheat bran to produce glucose syrup and functional cereal bars. *Bioprocess and Biosystems Engineering*, 47(7), 1081-1094. <https://doi.org/10.1007/s00449-024-03032-1>
- Figueroa, J. G., Borrás-Linares, I., Del Pino-García, R., Curiel, J. A., Lozano-Sánchez, J., & Segura-Carretero, A. (2021). Functional ingredient from avocado peel: Microwave-assisted extraction, characterization and potential applications for the food industry. *Food Chemistry*, 352, 129300. <https://doi.org/10.1016/j.foodchem.2021.129300>
- Flores, C. A., Ordóñez, A. L., Balanza, M. E., Morant, M. A., & Bocci, D. (2021). Factibilidad de recuperación de principios bioactivos a partir de residuos de la industrialización de tomate. *Revista de Ingeniería y Ciencias Aplicadas*, 1(1), Article 1.
- Grisales-Mejía, J. F., Cedeño-Fierro, V., Ortega, J. P., Torres-Castañeda, H. G., Andrade-Mahecha, M. M., Martínez-Correa, H. A., Álvarez-Rivera, G., Mendiola, J. A., Cifuentes, A., & Ibañez, E. (2024). Advanced NADES-based extraction processes for the recovery of phenolic compounds from Hass avocado residues: A sustainable valorization strategy. *Separation and Purification Technology*, 351, 128104. <https://doi.org/10.1016/j.seppur.2024.128104>
- Idris, S. A., Rosli, N. R., & Raja Aris, R. M. A. (2022). Supercritical carbon dioxide extraction of fatty acids compounds from tamarind seeds. *Materials Today: Proceedings*, 63, S462-S466. <https://doi.org/10.1016/j.matpr.2022.04.129>
- Jofre, V. P., Assof Roa, M. V., Vargas, E. V., & Fanzone, M. L. (2020). Determinación simultánea de carotenoides y compuestos fenólicos en subproductos agroalimentarios: Microextracción Sólido-Líquido asistida por ultrasonido optimizada por metodologías de diseño experimental. *Investigación, Ciencia y Universidad*, 4(5): 20-28.
- Jordan Suarez, O. B. (2019). Microencapsulación de extracto de hojas de guanábana (*Annona muricata* L.). Tesis. Universidad Nacional Agraria la Molina.
- Ku, C. Y. (2021). Extracción, caracterización y estabilidad de las antocianinas de cáscara de *Chrysophyllum cainito* L., cultivado en Yucatán. Tesis. Tecnológico Nacional de México.
- Leyva-López, N., Lizárraga-Velázquez, C. E., Hernández, C., & Sánchez-Gutiérrez, E. Y. (2020). Exploitation of agro-industrial waste as potential source of bioactive compounds for aquaculture. *Foods*, 9(7), Article 7. <https://doi.org/10.3390/foods9070843>
- López-Linares, J. C., Campillo, V., Coca, M., Lucas, S., & García-Cubero, M. T. (2021). Microwave-assisted deep eutectic solvent extraction of phenolic compounds from brewer's spent grain. *Journal of Chemical Technology and Biotechnology*, 96(2), 481-490. <https://doi.org/10.1002/jctb.6565>
- Marentes, N. A. M., Ramírez, E. J. H., & Díaz, M. P. T. (2019). Caracterización y estabilidad de un extracto rico en antocianinas a partir del epicarpio de gulupa (*Passiflora edulis* f. *Edulis* Sims). *Revista Colombiana de Química*, 48
- Martínez Rodríguez, M. (2021). Aprovechamiento de dos subproductos agroindustriales en el desarrollo de un snack rico en fibra dietaria. Tesis. Universidad Nacional de Colombia.
- Meral, R., Kose, Y. E., Ceylan, Z., & Cavidoglu, İ. (2022). Chapter 10. The potential use of agro-industrial by-products as sources of bioactive compounds: A nanotechnological approach. En Atta-Ur-Rahman (Ed.), *Studies in natural products chemistry*, 73, 435-466. <https://doi.org/10.1016/B978-0-323-91097-2.00014-5>
- Molina-Hernández, J. B., Martínez-Correa, H. A., & Andrade-Mahecha, M. M. (2019). Potencial Agroindustrial del epicarpio de Maracuyá como ingrediente alimenticio activo. *Información Tecnológica*, 30(2), 245-256. <https://doi.org/10.4067/S0718-07642019000200245>
- Monzón, L., Becerra, G., Aguirre, E., Rodríguez, G., & Villanueva, E. (2021). Ultrasound-assisted extraction of polyphenols from avocado residues: Modeling and optimization using response surface methodology and artificial neural networks. *Scientia Agropecuaria*, 12(1), 33-40. <https://doi.org/10.17268/sci.agropecu.2021.004>
- Navarro-Valdez, K., Capillo-Herrera, N., Calixto-Cotos, M. R., & Santisteban-Rojas, O. P. (2020). Extraction and microencapsulation of antioxidant compounds from *Oenocarpus bataua* Mart seed. *Scientia Agropecuaria*, 11(4), 547-554. <https://doi.org/10.17268/sci.agropecu.2020.04.10>
- Nemes, S. A., Szabo, K., & Vodnar, D. C. (2020). Applicability of agro-industrial By-Products in intelligent food packaging. *Coatings*, 10(6), Article 6. <https://doi.org/10.3390/coatings10060550>
- Ordóñez-Santos, L. E., Velasco-Arango, V. A., & Hleap-Zapata, J. I. (2022). Ultrasound-assisted extraction of total carotenoids in papaya epicarp and its application in Frankfurt sausage. *Ciência e Agrotecnologia*, 46, e006722. <https://doi.org/10.1590/1413-7054202246006722>
- Pacheco, M. T. (2019). *Obtención y caracterización de compuestos bioactivos procedentes de tubérculos andinos y de subproductos de la industria agroalimentaria* [Doctoral thesis]. Universidad Autónoma de Madrid.
- Pal, C. B. T., & Jadeja, G. C. (2020). Microwave-assisted extraction for recovery of polyphenolic antioxidants from ripe mango (*Mangifera indica* L.) peel using lactic acid/sodium acetate deep eutectic mixtures. *Food Science and Technology International*, 26(1), 78-92. <https://doi.org/10.1177/1082013219870010>
- Paz, S. L. (2022). Extracción de compuestos bioactivos de residuos de piña (*Ananas comosus*) usando fermentación en estado sólido. Universidad Nacional de Colombia. <https://repositorio.unal.edu.co/handle/unal/84174>
- Portillo-López, R., Morales-Contreras, B. E., Lozano-Guzmán, E., Basilio-Heredia, J., Muy-Rangel, M. D., Ochoa-Martínez, L. A., Rosas-Flores, W., & Morales-Castro, J. (2021). Vegetable oils as green solvents for carotenoid extraction from pumpkin (*Cucurbita argyrosperma* Huber) by-products: Optimization of extraction parameters. *Journal of Food Science*, 86(7), 3122-3136. <https://doi.org/10.1111/1750-3841.15815>
- Reguengo, L. M., Salgaço, M. K., Sivieri, K., & Maróstica Júnior, M. R. (2022). Agro-industrial by-products: Valuable sources of bioactive compounds. *Food Research International*, 152, 110871. <https://doi.org/10.1016/j.foodres.2021.110871>
- Robles-Botero, M. V., Ronquillo-de Jesús, E., Quiroz-Reyes, C. N., & Aguilar-Méndez, M. A. (2020). Caracterización e

- identificación de compuestos bioactivos con actividad antioxidante de la cáscara, pulpa y semilla del fruto de tejocote (*Crataegus mexicana*). *TIP Revista Especializada en Ciencias Químico-Biológicas*, 23. <https://doi.org/10.22201/fesz.23958723e.2020.0.233>
- Rodríguez, S. B. O. (2023). Extracción verde ultrasónica de un antioxidante de subproducto de granada (*Punica granatum* L.) que preserva las propiedades de calidad en hamburguesas de pollo. Tesis de Máster. Universidad Autónoma de Querétaro. México.
- Rojas, E. F., Hernández-Carrión, M., & Sánchez-Camargo, A. (2022). Valorización de un subproducto del procesamiento de café arábica variedad caturra rojo y amarillo: Caracterización proximal, extracción de carotenoides y aplicación en un alimento funcional. Trabajo de grado. Universidad de los Andes. Colombia.
- Rojas, T., Gómez, S., Gómez, S., Fuentes Campos, M. E., Contreras-López, E., & Muñoz Jáuregui, A. M. (2019). Extracción asistida por ultrasonido de compuestos fenólicos de la cáscara de sanky (*Corryocactus brevistylus*). *Revista de la Sociedad Química del Perú*, 85(2), 258-267.
- Rubiano-Charry, K. D., Ciro-Velásquez, H. J., & Aristizabal-Torres, I. D. (2019). Aprovechamiento de los subproductos del mango, como fuente de compuestos bioactivos, para la elaboración de rollos comestibles. *Revista U.D.C.A Actualidad & Divulgación Científica*, 22(2), Article 2. <https://doi.org/10.31910/rudca.v22.n2.2019.1078>
- Saini, A., Kumar, A., Panesar, P. S., & Thakur, A. (2022). Potential of deep eutectic solvents in the extraction of value-added compounds from agro-industrial by-products. *Applied Food Research*, 2(2), 100211. <https://doi.org/10.1016/j.afres.2022.100211>
- Sangwan, K., Garhwal, R., Mehra, R., Pal, Y., Bhardwaj, A., Nayan, V., Kumar, N., Kumar, S., Goksen, G., Lorenzo, J. M., & Kumar, H. (2023). Development and characterization of W/O/W double emulsion of watermelon rind powder. *LWT*, 182, 114848. <https://doi.org/10.1016/j.lwt.2023.114848>
- Sharma, R., & Ghoshal, G. (2020). Optimization of carotenoids production by *Rhodotorula mucilaginosa* (MTCC-1403) using agro-industrial waste in bioreactor: A statistical approach. *Biotechnology Reports*, 25, e00407. <https://doi.org/10.1016/j.btre.2019.e00407>
- Shirahigue, L. D., & Ceccato-Antonini, S. R. (2020). Agro-industrial wastes as sources of bioactive compounds for food and fermentation industries. *Ciência Rural*, 50(4), e20190857. <https://doi.org/10.1590/0103-8478cr20190857>
- Simitzis, P. E. (2018). Agro-industrial By-Products and their bioactive compounds—An ally against oxidative stress and skin aging. *Cosmetics*, 5(4), Article 4. <https://doi.org/10.3390/cosmetics5040058>
- Surco Laos, F., Ayquipa Paucar, H., Quispe Gamboa, W., García Ceccarelli, J., & Valle Campos, M. (2020). Determinación de polifenoles totales y actividad antioxidante de extracto de semillas de uvas residuos de la producción de Pisos. *Revista de la Sociedad Química del Perú*, 86(2), 123-131. <https://doi.org/10.37761/rsqp.v86i2.282>
- Tarazona-Díaz, M., Becerra, N., Piedra, J., Beltrán, R., Tarazona-Díaz, M., Becerra, N., Piedra, J., & Beltrán, R. (2020). Obtención de un colorante a partir de corteza de maracuyá con el uso de técnicas convencionales de extracción. *Revista U.D.C.A Actualidad & Divulgación Científica*, 23(1). <https://doi.org/10.31910/rudca.v23.n1.2020.1303>
- Torres-Mendoza, K. E., Lara-Tambaco, R. M., & León-Araujo, M. E. (2023). Obtención y caracterización de la pectina extraída de la cáscara de pepino (*Cucumis sativus* L.-Variedad Híbrido Dasher II) y validarla como producto gelificante en compotas. *Ibero-American Journal of Engineering & Technology Studies*, 3(1), Article 1. <https://doi.org/10.56183/iberotecs.v3i1.597>
- Torres-Valenzuela, L. S., Serna-Jiménez, J. A., Pinto, V., & Vargas, D. (2020). Evaluación de condiciones de extracción asistida por ultrasonido de compuestos bioactivos de cáscara de pitahaya amarilla. *Revista Lasallista de Investigación*, 17(1), 70-83. <https://doi.org/10.22507/rli.v17n1a6>
- Urrialde, R., Gómez-Cifuentes, A., Pintos, B., Gómez-Garay, M. A., Cifuentes, B., Urrialde, R., Gómez-Cifuentes, A., Pintos, B., Gómez-Garay, M. A., & Cifuentes, B. (2022). Compuestos bioactivos de origen vegetal: Desarrollo de nuevos alimentos. *Nutrición Hospitalaria*, 39, (SPE3), 8-11. <https://doi.org/10.20960/nh.04302>
- Vargas-Sánchez, R., Torrecano-Urrutia, G., Esqueda-Valle, M., Torres-Martínez, B., & Sánchez-Escalante, A. (2022). Fermentación de subproductos agroindustriales como estrategia para obtener aditivos para alimento de codorniz. *Abanico Veterinario*, 12. <https://doi.org/10.21929/abavet2022.17>
- Viana, H. N. A. C., Sganzerla, W. G., Castro, L. E. N., & Veeck, A. P. L. (2023). Characterization of baru (*Dipteryx alata* Vog.) and application of its agro-industrial by-product in the formulation of cookies. *Journal of Agriculture and Food Research*, 12, 100577. <https://doi.org/10.1016/j.jafr.2023.100577>
- Villamil-Galindo, E., Van de Velde, F., & Piagentini, A. M. (2021). Strawberry agro-industrial by-products as a source of bioactive compounds: Effect of cultivar on the phenolic profile and the antioxidant capacity. *Bioresources and Bioprocessing*, 8(1), 61. <https://doi.org/10.1186/s40643-021-00416-z>
- Vuksinic, E. (2019). Caracterización química y ambiental de efluentes agroindustriales y análisis de sus implicancias sobre el agua subterránea del Valle Central Antinaco-Los Colorados, Provincia de la Rioja. Tesis, Universidad Nacional de Luján.