



Changes in physical and chemical characteristics of fermented cocoa (*Theobroma cacao*) beans with manual and semi-mechanized transfer, between fermentation boxes

Cambios en la características físicas y químicas de granos de cacao (*Theobroma cacao*) fermentados con transferencia manual y semi-mecanizada, entre las cajas de fermentación

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Abstract

The aim of this study was to evaluate variation in the physical and chemical properties of fermented cocoa beans with cocoa beans transfer between wooden fermentation boxes manually (M) and semi-mechanized (SM) way. Mass temperature, moisture, pH, and total acidity of the cotyledon and pulp; the total polyphenol, anthocyanin, reducing sugar, theobromine, and caffeine content in fresh, fermented, and dried beans; and percentage of fermented beans and time required to move beans during fermentation were determined. The cocoa used grew in the Pachiza district of the San Martin region of Peru. Cocoa sampling was each 0, 48, 72, 96, 120, 144, and 168 h of fermentation. The cocoa mass temperature was highest with both removal systems after 96 h of fermentation. M cotyledon and pulp samples had the highest moisture content and titratable acidity, while cotyledon and pulp pH with both systems were statistically equal. In contrast, fermented beans had a higher polyphenol, anthocyanin, reducing sugar, theobromine, and caffeine content with SM. SM produced the greatest amount of fermentation (91.67%) and required the shortest amount of time to move beans (78.56 min). In conclusion, the system of fermentation of cocoa beans with SM was faster and produced fermented grains with high chemical quality.

Keywords: Fermented cocoa, transfer, manually, semi-mechanized.

1. Introduction

Cocoa postharvest handling is very important and determines the quality of the product on the market (Sandhya *et al.*, 2016). Fermentation of cocoa beans is the first step in the chocolate-making chain (De Melo Pereira *et al.*, 2013). Cocoa bean fermentation is very important and beneficial; microbial fermentation of cocoa removes mucilage (Hatmi *et al.*, 2015) and induces a set of internal biochemical reactions in the cotyledon that lead to modification of the chemical composition of cocoa beans and the formation of aromatic precursors. During fermentation, the microbial succession occurs by changes in temperature, pH and

availability of oxygen (Kongor *et al.*, 2016). As the aeration increases due to further cocoa pulp drainage and the temperature of the fermenting cocoa pulp-bean mass increases above 37 °C. The environmental conditions become favorable for the growth of acetic acid bacteria, which oxidize the ethanol produced by yeasts and lactic acid produced by lactic acid bacteria into acetic acid and subsequently overoxidize this acetic acid into carbon dioxide and water (Illegheems *et al.*, 2015). Hence, when and how fast the cocoa mass is turned up is important for achieving the desired amount of acetic acid fermentation; improper removal promotes lactic acid fermentation

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which ultimately affects the quality of commercial cocoa (Afoakwa *et al.*, 2011). In order to determine the effect of manually (M) and semi-mechanized (SM) of cocoa beans transfer during fermentation, we evaluated the temperature, moisture content, pH, and total acidity in the cotyledon and pulp, as well as quantified the total polyphenol, anthocyanin, reducing sugar, theobromine, and caffeine content in fresh, fermented, and dried cocoa beans. The aim of this study was to evaluate variation in the physical and chemical properties of cocoa bean fermentation with cocoa beans transfer manually (M) and semi-mechanized (SM) way in order to improve the conditions in the processing.

2. Materials and methods

2.1 Raw material

Cocoa beans: This research used Forastero hybrid and CCN-51 cocoa beans from the Pachiza district of Mariscal Caceres province in the San Martin region of Peru. Beans were grown at an altitude of 328 m to 07° 17 '49" south latitude and 76° 46 '17" north latitude. Farmers harvest cacao pods, break pods, get the mucilage along with grains. The local fermentation uses wooden boxes of 1 m³ capacity, standing cocoa beans in the box, where a spontaneous fermentation occurs with 500 kg, for 168 h at room temperature. Cocoa beans were transferred manual and semi-mechanized way, from one box to another, once per day, after the second day to obtain a uniform fermentation.

2.2 Analytical procedures

Temperature

The temperature measurements in the cocoa mass was in the center and on the environment, performed after 0, 48, 72, 96, 120, 144, and 168 h of fermentation.

pH

pH of the cocoa pulp and cotyledons were determined using a pH meter after 0, 48, 72, 96, 120, 144, and 168 h of fermentation according to Method 931.04 (AOAC, 1995).

Acidity: Total titratable acidity as acetic acid (g/100 g cocoa), in the cocoa pulp and cotyledons, was determined after 0, 48, 72, 96, 120, 144, and 168 h of fermentation using Method 942.15 (AOAC, 1995).

Moisture

Total moisture content (%) in the cocoa mass and surrounding environment was determined after 0, 48, 72, 96, 120, 144, and 168 h of fermentation using Method 931.04 (AOAC, 1995).

Polyphenols

Total polyphenol content was quantified using the Folin-Ciocalteu method in cocoa bean extracts after 0, 48, 72, 96, 120, 144, and 168 h of fermentation; results were expressed in gallic acid equivalents [mg GAE/g extract] (Symonowicz *et al.*, 2012; Sultana *et al.*, 2012).

Anthocyanins

Total anthocyanin content (mg cyanidin-3-glucoside/g cocoa) was measured by the pH differential method described by Symonowicz *et al.* (2012) after 0, 48, 72, 96, 120, 144, and 168 h of fermentation.

Reducing sugars

Total amount of unfermented (reducing) sugar (%) in cocoa beans was determined using dinitrosalicylic acid and quantified (mg sugar/g cocoa) from a standard curve plotted based on the absorbance at 510 nm (Miller, 1959) after 0, 48, 72, 96, 120, 144, and 168 h of fermentation.

Theobromine and caffeine

Analysis of total theobromine (g theobromine/g cocoa) and caffeine (g caffeine/g cocoa) content used a Shimadzu LC-10AT reverse-phase liquid chromatograph (VP Scientific, Columbia, MD 21046, U.S.A.). Consisting of a Shimadzu DGU-14A degasser, manual Rheodyne 7725i injector, management pack solvent with Shimadzu LC-10AT quaternary pump, Shimadzu CTO-10AS single-column oven, and Shimadzu UV-Vis SPD-10AV detector. A Shimadzu SCL-10AV interface to determine chromatographic peak purity, and identification and integration of peaks used Shimadzu CLASS-VP™ software version 6.13 SP2. Chromatographic separation was

completed using a C18 Ultra guard cartridge (20 mm x 4 mm; Code: 917450220; Restek, Bellefonte, PA 16823, United States with a C18 Ultra column (150 mm x 4.6 mm x 5 μ m; Code: 9174565; Restek, Bellefonte, PA 16823, United States. The column temperature was kept at 35 °C. Detection of theobromine and caffeine in fresh, fermented (0, 48, 72, 96, 120, 144, and 168 h), and dry beans with a 7% moisture content was performed at 210 nm. The mobile phase was a mixture of acetic acid (0.3%) and methanol (85:15) and samples solutions filtered with a nylon microfilter (0.2 μ m), the flow rate was 1 ml/min, and the sample volume was 20 μ l (Lo Coco *et al.*, 2007; Menguy *et al.*, 2009). The analysis was conducted in triplicates and the mean values reported.

Cutting test

This test was conducted to physically evaluate and determine the percentage of good quality, dried, fermented cocoa beans by separating them from defective beans (e.g., purple or partially violet); the sum of the number of fermented and defective beans represented 100% (National Federation of Cocoa - Cocoa National Fund, 2004).

Experimental design and statistical analysis

Our study had a completely randomized design and replicated thrice followed by Tukey's test to determine statistical differences using Centurion Statgraphics XV.V.15.2.06 software (De Mendiburu,

2007). A p-value < 0.05 was considered statistically significant. All values are presented as means (Me) \pm standard error of the mean (SEM) unless indicated otherwise.

3. Results and discussion

3.1. Cocoa mass and environment temperature

The highest mean temperature of the cocoa mass (47.7 °C) occurred after 96 h of fermentation for both M and SM (Table 1). However, cocoa mass temperatures were statistically different ($p \leq 0.05$) between the two transfer ways at other time points (Table 1). It reaffirms the results reported by Amores *et al.* (2009) who suggested that early on in the fermentation process cocoa mass temperatures normally vary between 45 and 50 °C. Schwan (1998) reported that when the cocoa mass temperature raises to about 50 °C, and the heat and acid result in chemical reactions in the beans known as curing, production of organic acids (oxalic, phosphoric, succinic, malic, and acetic acids).

The mean temperature of the cocoa mass at the end of fermentation (168 h) was 45.13 ± 2.34 °C with SM and 41.90 ± 0.60 °C with M. The mean cocoa mass temperature decreased after 96 h due to inactivation of predominant bacteria at temperatures >40 °C and embryonic death caused by acetic acid penetration into the bean, favoring development of chocolate flavor precursors (Kongor *et al.*, 2016).

Table 1

Mean temperature of the cocoa mass and surrounding environment during fermentation

Fermentation time (h)	Manually Cocoa Beans Transfer		Semi-mechanized Cocoa Beans Transfer	
	TCM (°C)	ET (°C)	TCM (°C)	ET (°C)
0	28.30 \pm 0.43 ^a	28.00 \pm 0.00	29.77 \pm 0.18 ^a	25.85 \pm 2.15
48	35.7 \pm 0.09 ^{ab}	24.90 \pm 0.10	39.80 \pm 3.60 ^{bc}	25.50 \pm 0.50
72	40.23 \pm 0.24 ^{bc}	25.00 \pm 0.30	44.47 \pm 2.10 ^{cd}	26.60 \pm 0.60
96	47.70 \pm 0.15 ^d	27.95 \pm 0.45	47.70 \pm 0.12 ^d	27.40 \pm 3.20
120	43.67 \pm 0.03 ^{cd}	27.90 \pm 1.20	45.47 \pm 0.94 ^{cd}	27.75 \pm 1.35
144	45.03 \pm 0.35 ^{cd}	27.10 \pm 0.30	45.73 \pm 0.43 ^{cd}	26.50 \pm 0.90
168	41.90 \pm 0.60 ^{cd}	26.05 \pm 0.25	45.13 \pm 2.34 ^{cd}	25.75 \pm 0.55

TCM: Cocoa mass temperature; ET: Environment temperature. Values in the same column with the same superscript letters are statistically equal. Values represent the Me \pm SEM; n = 3; $p \leq 0.05$, Tukey's test.

3.2. Moisture variation in cocoa beans over time

The mean moisture content of cocoa beans was statistically different ($p \leq 0.05$) throughout the fermentation process (Figure 1) and may be due to the variety and maturity of the fruit. The highest mean moisture content existed in fresh beans (0 h) with both M ($51.89 \pm 1.74\%$) and SM ($51.91 \pm 0.74\%$). After 168 hours of fermentation, the moisture content fell to $46.33 \pm 0.60\%$ with M and $42.77 \pm 1.90\%$ with SM ($p > 0.05$). Similar to these results, Rodriguez *et al.* (2012) reported 43.7% moisture in fermented beans. Furthermore, stated that fermentation of cocoa bean pulp by microbial action causes cell rupture and release of intracellular juices, thereby reducing the amount of moisture retained by beans.

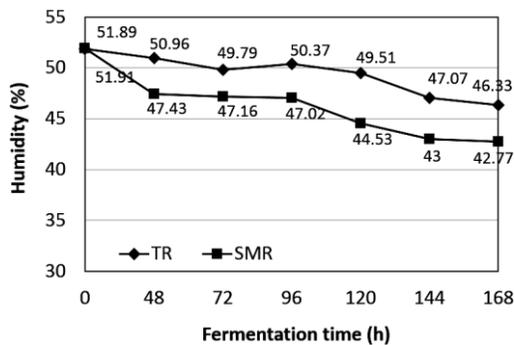


Figure 1. Moisture content in fermented cocoa beans over time with manually (M) and semi-mechanical (SM) cocoa beans transfer.

Statistical analysis revealed significant differences ($p \leq 0.05$) between the pH of the pulp and cotyledon of cocoa beans with both M and SM throughout the ferment-

ation process (Table 2). However, the pH of the pulp and cotyledon were statistically equal by the end of fermentation (168 h). The pH of the cotyledon decreased consistently from 6.99 ± 0.20 in fresh beans (0 h) to 4.76 ± 0.03 in fully fermented beans (168 h) with M. Similarly, Graziani de Fariñas *et al.* (2002) reported a fermented cocoa bean cotyledon pH of 4.75. Amores *et al.* (2009) suggested that acetic acid infiltrates the cotyledon and lowers the pH from 6.4 to 4.5 during fermentation at temperatures $>45\text{ }^\circ\text{C}$. This acidification disintegrates compartments of the cell and eventually leads to cellular death. The cocoa bean pulp is permeable to acetic acid, which then passes into the cotyledon after three days, killing the embryo and lowering the pH to 4.8. Furthermore, Afoakwa *et al.* (2008) postulated that poorly fermented cocoa has a pH of 5.5-5.8 while properly fermented cocoa has a pH of 4.7-5.2.

3.3. Acidity of cocoa bean pulp and cotyledons

Statistical analysis revealed significant differences ($p \leq 0.05$) between the total titratable acidity of the pulp and cotyledon of cocoa beans with both M and SM throughout the fermentation process (Table 3). Cotyledon acidity increased to 2.88 ± 0.18 as acetic acid (g/100 g cocoa), with M by the end of fermentation (168 h). Rivera *et al.* (2012) indicated that acids produced by microorganisms during fermentation cause an increase in acidity and consequent decrease in pH.

Table 2
Variation pH of cocoa beans during fermentation

Fermentation time (h)	Manually Cocoa Beans Transfer		Semi-mechanized Cocoa Beans Transfer	
	Pulp	Cotyledon	Pulp	Cotyledon
0	4.21 ± 0.02^a	6.99 ± 0.20^d	4.20 ± 0.03^a	7.08 ± 0.09^d
48	4.39 ± 0.05^{ab}	5.98 ± 0.02^c	4.50 ± 0.04^{bc}	5.87 ± 0.43^{bc}
72	4.63 ± 0.08^{cd}	5.37 ± 0.11^{abc}	4.67 ± 0.07^{cde}	5.07 ± 0.40^{ab}
96	4.77 ± 0.05^{de}	5.05 ± 0.03^{ab}	4.79 ± 0.05^{de}	5.00 ± 0.02^{ab}
120	4.80 ± 0.03^{de}	4.91 ± 0.06^a	4.85 ± 0.03^e	4.93 ± 0.02^a
144	4.79 ± 0.05^{de}	4.81 ± 0.02^a	4.86 ± 0.02^e	4.86 ± 0.01^a
168	4.81 ± 0.01^{de}	4.76 ± 0.03^a	4.83 ± 0.02^{de}	4.79 ± 0.01^a

Values in the same column with the same superscript letters are statistically equal. Values represent the Me \pm SEM; n = 3; $p \leq 0.05$, Tukey's test.

Table 3

Variation of acidity of cocoa beans during fermentation

Fermentation time (h)	Manually Cocoa Beans Transfer		Semi-mechanized Cocoa Beans Transfer	
	Pulp	Cotyledon	Pulp	Cotyledon
0	0.96 ± 0.19 ^{ab}	0.77 ± 0.19 ^a	0.77 ± 0.19 ^a	0.58 ± 0.00 ^a
48	1.44 ± 0.18 ^{bc}	1.08 ± 0.00 ^{ab}	1.62 ± 0.00 ^{bcd}	1.08 ± 0.00 ^{ab}
72	1.98 ± 0.18 ^{cde}	1.62 ± 0.00 ^{bc}	2.16 ± 0.00 ^{def}	1.62 ± 0.00 ^{bc}
96	2.16 ± 0.00 ^{def}	1.98 ± 0.18 ^{cd}	2.16 ± 0.00 ^{def}	2.16 ± 0.00 ^{cde}
120	2.52 ± 0.18 ^{efg}	2.16 ± 0.00 ^{cde}	2.52 ± 0.18 ^{efg}	2.34 ± 0.18 ^{cde}
144	2.70 ± 0.00 ^{fg}	2.52 ± 0.18 ^{de}	2.70 ± 0.00 ^{fg}	2.16 ± 0.31 ^{cde}
168	3.06 ± 0.18 ^g	2.88 ± 0.18 ^e	2.70 ± 0.00 ^{fg}	2.34 ± 0.18 ^{cde}

Values in the same column with the same superscript letters are statistically equal. Values represent the Me ± SEM; n = 3; p ≤ 0.05, Tukey's test.

The acidity of the fresh cotyledon (0 h) was lowest with SM (0.58 ± 0.00 as acetic acid g/100 g cocoa) and increased to a peak of 2.34 ± 0.18 after 120 h of fermentation; upon completion of fermentation (168 h), the acidity was 2.34 ± 0.18 as acetic acid g/100 g cocoa. These results were similar to those reported an acidity level of 2.37 for fermented cocoa cotyledons. The total concentration of acids was significantly higher than all the other chemical groups. The total acids concentration increased significantly at the beginning of fermentation (2 and 4 days) (Rodriguez *et al.*, 2012).

3.4. Total polyphenol content in cocoa beans during fermentation

The results showed statistically significant differences ($p \leq 0.05$) in total polyphenol content between M and SM samples throughout fermentation (Figure 2). The highest total polyphenol content existed in fresh (0 h) cocoa beans (~7.0 g GAE/100 g cocoa) with either M or SM. Niemenak *et al.* (2006) indicated that different polyphenol contents in cocoa clones are partly due to genetic characteristics (i.e., variety) and environmental variables, such as growth conditions, light intensity, humidity, temperature, fertilizer use, health, stress, etc.

Figure 2 shows that the total polyphenol content in cocoa beans decreased throughout the fermentation process, resulting in 5.88 ± 0.56 g GAE/100 g cocoa with SM and 5.05 ± 0.02 g GAE/100 g cocoa with M after 168 h. These results were similar to those reported by Zapata *et*

al. (2013) in fermented, dried Trinitario cocoa (5.02 g GAE/100 g cocoa), also showed a decrease in polyphenol content between fresh and fermented beans. Niemenak *et al.* (2006) suggested this decrease is due to the spread of polyphenols outside the cotyledon during fermentation. Othman *et al.* (2007) suggested that polyphenol content differs by bean variety, the degree of fermentation, and different processing parameters.

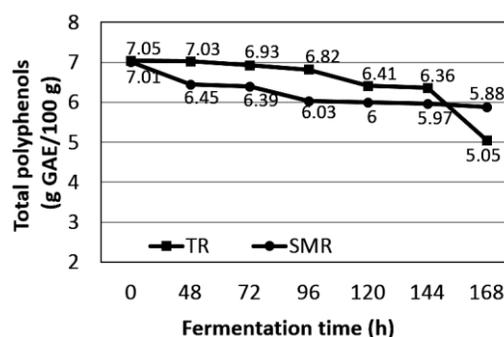


Figure 2. Total polyphenol content in cocoa beans during fermentation with manually (M) and semi-mechanized (SM) cocoa beans transfer.

3.5. Anthocyanin content in cocoa beans during fermentation

The results showed a statistically significant difference ($p \leq 0.05$) in anthocyanin content among M and SM samples throughout fermentation (Figure 3). Fresh cocoa beans (0 h) had the highest anthocyanin content with both SM and M (15.59 ± 0.44 and 13.26 ± 0.96 mg cyanidin-3-glucoside/g cocoa, respectively).

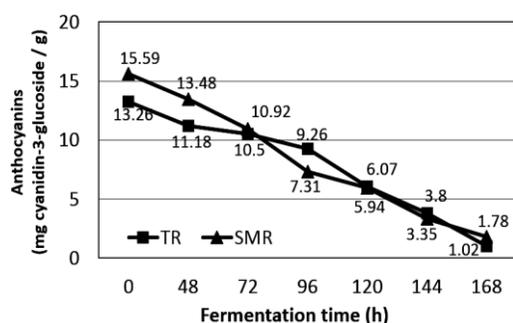


Figure 3. Anthocyanin content in cocoa beans during fermentation with manually (M) and semi-mechanized (SM) cocoa beans transfer.

Niemenak *et al.* (2006) reported the average value of total anthocyanin in freshly harvested, fermented beans from nine cocoa genotypes were 6587.5, 8239 and 6720 mg/kg dry matter respectively, obviously lower than current results. The cotyledon staining is a typical genetic characteristic associated with the cocoa variety and can vary from white (Creole) to highly pigmented (Forastero), with different shades and color distributions. Exists procyanidin in various proportions depending on the cocoa cultivar, which explains the existing empirical associations between color intensity and bitterness/astringency of pigmented beans, as well as differences in anthocyanin content between studies.

Figure 3 shows a rapid decrease in total anthocyanin content with both M and SM, resulting in 1.78 ± 0.71 and 1.02 ± 0.21 mg cyanidin-3-glucoside/g cocoa, respectively, after 168 h of fermentation. These results were similar to those found by Zapata *et al.* (2013) who reported 1.05 ± 0.045 mg cyanidin-3-glucoside/g of fermented Trinitario cocoa beans. The Forastero and Criollo cocoa beans undergo changes in chemical components during fermentation, demonstrated that anthocyanins undergo enzymatic hydrolysis by the action of yeast during fermentation while some alkaloids and polyphenols are lost by oxidation. During fermentation, the anthocyanin content decreased by 92% with M and 88% with SM compared to fresh beans (0 h).

3.6. Reducing sugar content in cocoa beans during fermentation

Statistical analysis showed significant differences ($p \leq 0.05$) in the reducing sugar content of cocoa beans during fermentation (Figure 4). The reducing sugar content was highest in fresh beans (0 h) with M (3.83%) and SM (3.41%). Both results were higher than those reported by Graziani *et al.* (2003) who found 3.39 % reducing sugar content in fresh beans. These observed differences may be due to factors such as bean variety and maturity of fruit.

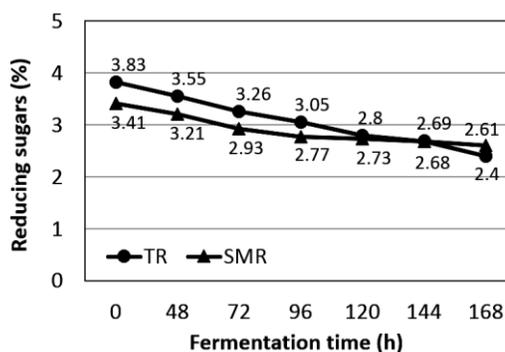


Figure 4. Reducing sugar content in cocoa beans during fermentation with manually (M) and semi-mechanized (SM) cocoa beans transfer.

The amount of reducing sugars decreased for both M and SM throughout cocoa beans fermentation process, with 2.61% remaining with SM and 2.40% with M after 168 h (Figure 4). Continuous chemical and biochemical changes occur during fermentation. For example, sugars are reduced by participating in no enzymatic browning reactions, which are favored by high temperatures (30-50 °C). When drying proceeds slowly, the term water activity remains high for a longer period, which is optimal for Maillard chemistry (Cros y Jeanjean, 1995). Under these conditions, Maillard reactions thrive along with formation of the volatile fraction (Afoakawa, 2010).

3.7. Theobromine content in cocoa beans during fermentation

The results revealed statistically significant differences ($p \leq 0.05$) in theobromine

content throughout the process of fermentation for both types of systems (Fig. 5). The highest theobromine content was found in fresh beans (0 h) with both SM and M (7.78 ± 0.23 and 6.93 ± 0.36 g theobromine/100 g cocoa, respectively). At the end of the fermentation process (168 h), theobromine concentrations decreased to 5.78 ± 0.19 and 5.21 ± 0.03 g theobromine/100 g cocoa with SM and M, respectively (Figure 5). Although Rodriguez *et al.* (2012) also reported a decrease after fermentation, the theobromine content (2.93 g theobromine/100 g fermented beans) was much lower than found herein. It is known that during the post-harvest cacao pods, fermenting and drying cocoa beans decreases the content of theobromine while the chemical quality improves with the reduction of alkaloids and polyphenols, which impacts the organoleptic characteristics of cocoa (Rivera *et al.*, 2012).

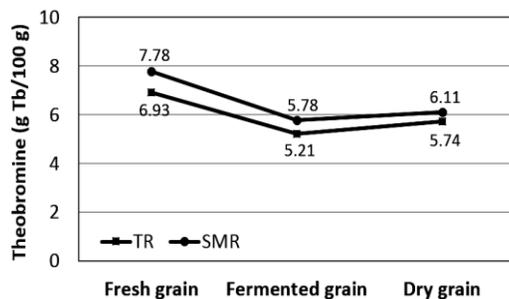


Figure 5. Theobromine content in fresh, fermented, and dried cocoa beans with manually (M) and semi-mechanized (SM) cocoa beans transfer.

3.8. Caffeine content in cocoa beans during fermentation

Statistical analysis revealed significant differences ($p \leq 0.05$) in fresh, fermented, and dried cocoa beans with both types of cocoa beans transfer (Figure 6). A higher caffeine content was found in fresh cocoa beans and varied between 0.39 ± 0.02 and 0.51 ± 0.02 g caffeine/100 g cocoa with SM and M, respectively. These results were similar to those reported by Nazaruddin *et al.* (2006) in fermented beans (0.40 g caffeine/100 g cocoa). The

decrease in alkaloid concentration is likely due to their diffusion out of the bean when the seed dies. Interestingly, the caffeine content with M and SM fell by 10.26% and 23.53% respectively, with fermentation.

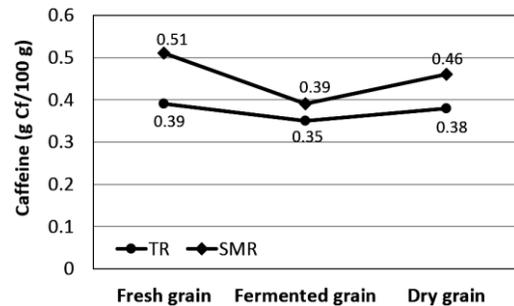


Figure 6. Variation of the caffeine content in fresh, fermented, and dried cocoa beans with manually (M) and semi-mechanized (SM) cocoa beans transfer.

3.9. Ratio of theobromine to caffeine in fermented cocoa beans

The ratio of theobromine:caffeine (Tb:Cf) with M and SM was 17.76 and 15.25, respectively, which were higher than that reported by Zambrano *et al.* (2010). In this regard suggested a Tb:Cf ratio >8 corresponds to the Foreigner variety; therefore, the Pachiza district of Peru has geographical areas that produce Foreigner cocoa.

3.10. Percentage of fermentation in dried cocoa beans

We found a significant difference ($p \leq 0.05$) between the types of cocoa beans transfer used during the fermentation process (Table 4). The highest percentage of fermented beans was $91.67 \pm 0.78\%$ with SM and $87.30 \pm 1.00\%$ with M. Furthermore, Alvarez *et al.* (2010) reported that a decrease in cocoa pulp acidity increases the proportion of brown beans. Development of brown pigments is an important stage during the drying process and is caused by the enzymatic oxidation of polyphenols, such as leucocyanidin and epicatechin, by polyphenol oxidase in the presence of oxygen and subsequent condensation of proteins (Ortiz de Bertorelli *et al.*, 2009).

Table 4

Percentage of fermentation in dried cocoa beans with manually and semi-mechanized cocoa beans transfer

Cocoa beans transfer	Fermentation (%)
Manually	87.30 ± 1.00 ^a
Semi-mechanized	91.67 ± 0.78 ^{b*}

Values with the same superscript letters are statistically equal. * significant. Values represent the Me ± SEM, n = 3, p ≤ 0.05, Tukey's test.

3.11. Time spent on manually (M) and semi-mechanized (SM) cocoa beans transfer

The time taken to move cocoa beans was statistically different ($p \leq 0.05$) between M and SM samples (Figure 7). Cocoa beans transfer at 168 h of fermentation took the longest amount of time using either SM (18.21 min) or M (17.22 min). It is also important to note that the total removal time increased by adding the time spent moving beans to dryers. Overall, employees spent less time with SM (78.56 min) than M (96.24 min) of cocoa beans (Figure 7). During the fermentation process, beans undergo a series of biochemical changes, which favor SM. The time difference between cocoa beans transfer methods was 17.68 min, this time difference between the methods is because the manual transfer of cocoa beans is done with more operator movements.

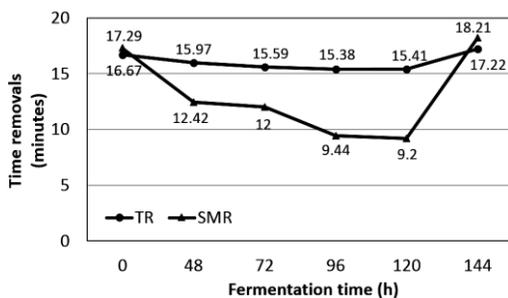


Figure 7. Cocoa beans removal time with manually (M) and semi-mechanized (SM) cocoa beans transfer.

4. Conclusions

Herein, the maximum fermentation temperature was 47.70 ± 0.12 °C with SM and 47.70 ± 0.15 °C with M after 96 h of

fermentation. The moisture content after 168 h of fermentation was $46.33 \pm 0.60\%$ with M and $42.77 \pm 1.90\%$ with SM. The cotyledon pH at the end of fermentation (168 h) was 4.76 ± 0.03 with M and 4.79 ± 0.01 with SM. The total titratable acidity was 2.88 ± 0.18 and 2.34 ± 0.18 as acetic acid (g/100 g cocoa), with M and SM, respectively. The highest total polyphenol content was 5.88 ± 0.18 g it is also important to note that the total removal time increased after adding the time spent moving beans to dryers AGE/100 g fermented cocoa beans with SM, while the lowest was 5.05 ± 0.02 g AGE/100 g cocoa with M. The highest anthocyanin content measured was 1.78 ± 0.71 mg cyanidin-3-glucoside/g fermented cocoa beans with SM, while the lowest was 1.02 ± 0.21 mg cyanidin-3-glucoside/g cocoa with M. The highest reducing sugar content was found in fermented cocoa beans with SM ($2.61 \pm 0.02\%$), while the lowest was associated with M ($2.40 \pm 0.16\%$). The theobromine content of dry beans was 5.74 ± 0.02 and 6.11 ± 0.14 g theobromine/100 g cocoa with M and SM, respectively, while the caffeine content in dry beans was 0.38 ± 0.01 and 0.46 ± 0.03 g caffeine/100 g cocoa, respectively. The ratio of Tb:Cf found suggests that the Pachiza district of Peru has Forastero cocoa plants growing in some areas. Overall, SM was found to produce the greatest amount of fermentation (91.67%) and required the least amount of employee time to move beans (78.56 min).

Acknowledgments

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