

## TRABAJOS ORIGINALES

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## Tabanidae (Diptera) del Perú: lista actualizada y descripción de tres nuevas especies

### Tabanidae (Diptera) of Peru: checklist update and description of three new species

#### Abstract

We provide an update to the list of the species of Tabanidae known from Peru, along with descriptions of three new species: *Diachlorus tenuimaculatus* n. sp., *Stenotabanus (Stenotabanus) carrascoi* n. sp. and *Stenotabanus (Stenotabanus) chainei* n. sp., bringing to 233 the species of Tabanidae now known from Peru.

#### Resumen

Se realizó una actualización de la lista de especies de Tabanidae del Perú y se describen tres nuevas especies, *Diachlorus tenuimaculatus* sp. n., *Stenotabanus (Stenotabanus) carrascoi* sp. n., y *Stenotabanus (Stenotabanus) chainei* sp. n., con ellas, suman 233 especies de Tabanidae registradas para Perú.

#### Keywords:

horseflies; taxonomy; Diachlorini; Neotropics; new species.

#### Palabras clave:

tábanos; taxonomía; Diachlorini; región Neotropical; nuevas especies.

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*Stenotabanus carrascoi* Henriques & Krolow, 2020

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#### Acto nomenclatural/nomenclatural act:

*Stenotabanus chainei* Henriques & Krolow, 2020

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## Introduction

Tabanidae (Diptera) have a worldwide distribution, currently with more than 4400 valid species. The greatest species richness is in the Neotropical region, with more than 1200 species — almost 30% of the world fauna (Henriques *et al.* 2012; Krolow *et al.* 2017). In an important contribution to knowledge of the tabanid fauna of Peru, Wilkerson and Fairchild (1985) tabulated the then-known fauna of 228 species (including subspecies and varieties). Several new species in the Peruvian fauna have been described since 1985, but there has been no subsequent, comprehensive update of the Peruvian fauna, either in list or catalog form.

Encouraged by the study of a few hundred specimens collected in Peru by Dr. José A. Rafael and his team, we here update and revise the Wilkerson and Fairchild (1985) checklist. This update includes species that Wilkerson and Fairchild (1985) had overlooked; adds species described after 1985; and deletes species that lack confirmation. As well, we describe three new species from Peru.

## Material and methods

The study material is derived from an insect-collecting trip to Peru by Dr. José A. Rafael and his team in August 2012. Four sites were sampled using a Malaise trap — a site in Madre de Dios and three sites in Cuzco (see the Appendix 1 for precise locality information). Specimens were examined and digitally photographed with a Leica M205 C stereomicroscope equipped with a Leica DFC 295 camera. The software used for digital-image processing was Leica Application Suite LAS V3.6. Morphological terminology follows Cumming & Wood (2017).

Following is the list of Institutions cited below, and their acronyms: Coleção de Entomologia da Universidade Federal do Tocantins, Porto Nacional, Brazil (CEUFT), Instituto Nacional de Pesquisas da Amazônia, Manaus, Brazil (INPA), Museu Paraense Emílio Goeldi, Belém, Brazil (MPEG), Museo de Historia Natural de la Universidad Nacional Mayor de San Marcos, Lima, Peru (MUSM), Museu de Zoologia da Universidade de São Paulo, São Paulo, Brazil (MZUSP).

**Table 1.** Updates to the Peru checklist of Wilkerson and Fairchild (1985).

Species	Record from Peru	Remarks
<i>Esenbeckia (Esenbeckia) perspicua</i> Wilkerson & Fairchild, 1983	Loreto: Iquitos	Henriques (2016)
<i>Esenbeckia (Esenbeckia) peruviana</i> Burger, 1999	Madre de Dios: Manu	described after 1985
<i>Fidena (Fidena) brevistria</i> (Lutz), 1909	not specified	Fairchild and Burger (1994)
<i>Scione flavohirta</i> Ricardo, 1902	not specified	Coscarón (2000)
<i>Betrequia ocellata</i> Oldroyd, 1970	Madre de Dios: Mazuko	new record
<i>Chrysops laetus</i> Fabricius, 1805	Marañón river	Henriques (2016)
<i>Chrysops rossi</i> Philip, 1960	Huánuco: Tingo María	record omitted from original description
<i>Acanthocera (Acanthocera) fairchildi</i> Henriques & Rafael, 1992	Madre de Dios: Puerto Maldonado	described after 1985
<i>Acanthocera (Acanthocera) gorayebi</i> Henriques & Rafael, 1992	Coronel Portillo: Pucallpa; Madre de Dios: Avispas	described after 1985
<i>Catachlorops (Amphichlorops) striatus</i> Burger, 1999	Pasco: Pan de Azúcar	described after 1985
<i>Catachlorops (Catachlorops) fumipennis</i> Kröber, 1931	lower Tapiche river	Henriques (2016)
<i>Chlorotabanus leuconotus</i> Krolow & Henriques, 2010	Madre de Dios: Manu	described after 1985
<i>Diachlorus anduzei</i> Stone, 1944	middle Ucayali river	Henriques (2016)
<i>Diachlorus tenuimaculatus</i> n. sp.	Cuzco: Quincemil; Madre de Dios: Mazuko	described here
<i>Diachlorus trevori</i> Wilkerson & Fairchild, 1982	Madre de Dios, Mazuko	new record
<i>Dichelacera (Desmatochelacera) albitibialis</i> Burger, 1999	Loreto: Boquerón; Pasco: Chontilla; Huánuco: Cachicoto	described after 1985
<i>Dichelacera (Dichelacera) rubrofemorata</i> Burger, 1999	Amazon river	described after 1985
<i>Di cladocera argentomacula</i> Wilkerson, 1979.	not specified	Fairchild and Burger (1994) as <i>argenteomaculata</i>
<i>Di cladocera fairchildi</i> Goodwin, 1999	Huánuco: Zapatagocha above Acomayo	described after 1985
<i>Stenotabanus (Stenotabanus) albiscutellatus</i> Chainey, 1999	Madre de Dios: Avispas, Loromayo	described after 1985
<i>Stenotabanus (Stenotabanus) carrascoi</i> n. sp.	Cuzco: Quincemil	described here
<i>Stenotabanus (Stenotabanus) chaineyi</i> n. sp.	Cuzco: Quincemil; Madre de Dios: Mazuko	described here
<i>Stenotabanus (Stenotabanus) nigriscapus</i> Chainey & Hall, 1999	Huánuco: Tingo María; Madre de Dios: Manu, Avispas	described after 1985
<i>Stenotabanus (Stenotabanus) penai</i> Chainey, 1999	Madre de Dios: Manu, Avispas	described after 1985
<i>Stypommisa anoriensis</i> Fairchild & Wilkerson, 1986	not specified	Fairchild and Burger (1994)
<i>Stypommisa furva</i> (Hine), 1920	Huánuco: Las Palmas	record omitted from Philip (1961), as <i>Stenotabanus</i>
<i>Tabanus fortis</i> Fairchild, 1961	not specified	Fairchild and Burger (1994)
<i>Tabanus noncallosus</i> Carmo & Henriques, 2019	Madre de Dios	described after 1985
<i>Tabanus xerodes</i> Philip, 1967	southern Peru	Fairchild and Burger (1994)

## Results and discussion

About 1000 specimens of 40 species were collected by Rafael and team. For details of their collection data and a species list see the Appendix 1.

Table 1 shows updated information for some species on the Wilkerson & Fairchild (1985) list and adds new records, including three new species, described below. This table increases the information in a previous catalog (Coscarón and Papavero 2009), which, except for the misidentification of *Stenotabanus taeniotes* (Wiedemann), did not include the records of Wilkerson & Fairchild (1985).

Table 2 shows the species that should be deleted from the Wilkerson and Fairchild (1985) list and provides a detailed rationale for their removal.

Wilkerson and Fairchild (1985) reported 228 species from Peru. As a result of additions and deletions, the new list of tabanid flies from Peru now numbers 233 species and three subspecies (see Appendix 2).

Turcatel (2019) provided a revision on the genus *Rhabdotylus*, listing Peru in the geographic distribution of *R. venenatum* (Osten Sacken), though it seems that she did not review material from this country. This species is not in the Wilkerson and Fairchild's checklist, besides the catalogue of Neotropical Diptera (Coscarón & Papavero 2009) does not include Peru as a known distribution for the species. We could not be able to track this record, therefore, not included this species in the list of Peruvian tabanids.

**Table 2.** Species of Tabanidae from the checklist of Wilkerson and Fairchild (1985) [= "W&F (85)"] not validated by this study.

Species	Rationale
<i>Esenbeckia (Esenbeckia) osornoi guianense</i> Fairchild, 1942	Now a synonym of <i>E. (E.) osornoi</i> Fairchild, 1942
<i>Esenbeckia (Proboscoides) ecuadorensis</i> Lutz and Castro, 1935	In W&F (85): "the specimens determined for Pechuman by Fairchild, are probably <i>E. (P.) suturalis</i> Rondani". Record not confirmed in subsequent catalogues
<i>Fidena (Fidena) howardi</i> Fairchild, 1941	In W&F (85): "specimen studied by Fairchild from Canadian National Collection, locality doubtful as it is known elsewhere only from Panama and Costa Rica". Record not confirmed in subsequent catalogues
<i>Fidena (Laphriomyia) rufopilosa</i> (Ricardo), 1900, as <i>F. (L.) mirabilis rufopilosus</i>	In W&F (85): "this identification not confirmed". Record not confirmed in subsequent catalogues
<i>Oscra rufa</i> (Macquart), 1838, as <i>Scaptia (Scaptia)</i>	In W&F (85): "Probably in error, all other records from Chile". Record not confirmed in subsequent catalogues
<i>Parosca albifrons</i> (Macquart), 1838, as <i>Scaptia (Pseudoscione)</i>	Recorded by Kröber (1951), but in W&F(85): "probably in error, no members of this subgenus known from Peru"
<i>Parosca latipalpis</i> (Macquart), 1850, as <i>Scaptia (Pseudoscione)</i>	Recorded by Soukup (1945), but in W&F(85): "probably in error, no members of this subgenus known from Peru"
<i>Scione incompleta</i> (Macquart), 1846	In W&F (85): "This species probably = <i>S. incompleta</i> of Kröber (1930b), not Macquart, which = <i>S. acer</i> Philip". Record not confirmed in subsequent catalogues
<i>Chrysops lutzi</i> Kröber, 1925	In W&F (85): "Peru" is given in original description but we doubt that it occurs in Peru since it is otherwise restricted to SE Brazil (Fairchild 1966)". Record not confirmed in subsequent catalogues
<i>Acanthocera (Acanthocera) bequaerti</i> Fairchild & Aitken, 1960	In the catalog of Coscarón and Papavero (2009) the species is restricted to Trinidad and Suriname. Probably these Peruvian specimens are of the species <i>A. fairchildi</i> Henriques & Rafael, 1992
<i>Acanthocera (Acanthocera) formosa</i> Kröber, 1930	Now a synonym of <i>A. marginalis</i> Walker, 1854
<i>Dasybasis lauta</i> var. <i>sublauta</i> Coscarón & Philip, 1967	Now a synonym of <i>Montismyia lauta</i> (Hine), 1920
<i>Dichelacera (Dichelacera) fuscipes</i> Lutz, 1915	In W&F (85): "Peru (Munich, possibly mislabelled)". Record not confirmed in subsequent catalogues
<i>Phaotabanus limpidadex</i> (Wiedemann), 1828	In W&F (85): "record of Kröber (1930a), but probably error". Probably is <i>P. phaeopterus</i> Fairchild, 1964. Record not confirmed in subsequent catalogues
<i>Stenotabanus (Stenotabanus) obscurus flavofemoratus</i> Kröber, 1929	Now a synonym of <i>S. obscurus</i> Kröber, 1929
<i>Stenotabanus (Stenotabanus) pallidicornis</i> Kröber, 1929	Now a synonym of <i>S. peruviansis</i> Kröber, 1929
<i>Stenotabanus (Stenotabanus) taeniotes</i> (Wiedemann), 1828	Recorded by Philip (1960). Misidentification, = <i>S. peruviansis</i> Kröber, 1929 (see Chainey et al. 1999)
<i>Tabanus albocirculus</i> Hine, 1907	In W&F (85): "Madre de Dios, Avispas (Canadian National Collection). Determination not certain, may be <i>T. rubripes</i> Macquart or <i>T. antarcticus</i> Linn". Record not confirmed in subsequent catalogues
<i>Tabanus flavicorpus</i> Philip, 1960	Now a synonymous of <i>T. fumomarginatus</i> Hine, 1920
<i>Tabanus fumatipennis</i> Kröber, 1933	Now a synonymous of <i>T. trivittatus</i> Fabricius, 1805
<i>Tabanus miles</i> Wiedemann, 1828	This species has been recorded to southeastern Brazil. The record of Soukup (1945) probably is wrong. Record not confirmed in subsequent catalogues
<i>Tabanus monotaxis</i> Philip, 1967	Record was not confirmed in subsequent catalogues
<i>Tabanus occidentalis</i> var. <i>modestus</i> Wiedemann, 1828 and var. <i>dorsovittatus</i> Macquart, 1955	Now a synonymous of <i>T. occidentalis</i> Linnaeus
<i>Tabanus peruvianus</i> Macquart, 1848	In W&F (85) "Peru" (Soukup 1945). Status of type not certain (BMNH)". See Fairchild and León (1986: 118). Record not confirmed in subsequent catalogues

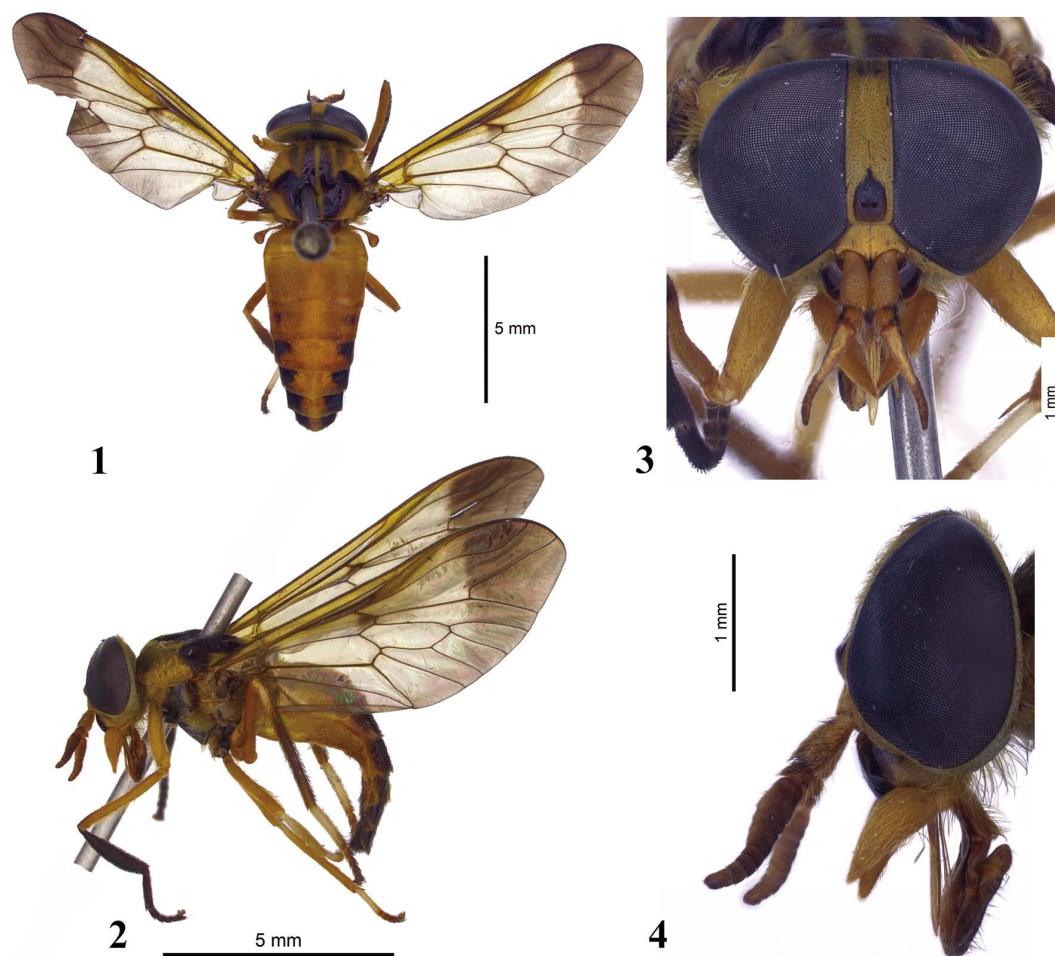
*Diachlorus tenuimaculatus* n. sp.

(Figs. 1 – 4)

A yellow species; scutum yellow with broad median shiny black spot reaching to posterior margin and laterally to wing base; the dark anterior-lateral spot is obscured. Wing with distinct brown apical patch that extends along the posterior margin until the anal lobe. Yellow abdomen with tergites 4–7 with black patches laterally.

**Female.** Length of holotype 8.5 mm (variation of paratypes 7.7 – 10.2 mm), wing 8.3 mm. Eye glabrous; the eye pattern is very similar to the pattern in *D. curvipes* (Fabricius) and *D. varipes* (Rondani), as illustrated in Lutz (1913), the second as *D. conspicuus* Lutz. Frons yellow pruinose with yellow hairs, black on vertex (divergence index = 1; frontal index = 4). Basal callus brown to dark-brown, narrower than frons, usually higher than wide. Subcallus, parafacial, gena, laterals of clypeus and palpus predominantly yellow pruinose with yellow hairs. Clypeus, except laterals, dark-brown shiny. Scape antennal yellow with yellow hairs dorsally and ventrally, black hairs laterally. Flagellum yellow-brown tomentose. First flagellomere yellow. Prementum and labella membranous and yellowish.

Prothorax and katatergite whitish. Scutum yellow with yellow hairs and pruinescence; in the middle a large black bare spot with a conspicuous yellow median stripe. The dark anterolateral spots may be inconspicuous by the pale pruinescence. Scutellum brown to black at the base, paler to the posterior margin. Notopleuron yellow with yellow hairs. Anepimeron and upper half of katepisternum yellowish; anepisternum and lower half of katepisternum blackish with the characteristic pearly pruinescence of the genus *Diachlorus*. Coxae and femora yellow with yellow hairs, except by black hairs at apex of fore femur. Fore tibia brown to black with black hairs, brown hairs ventrally (internally). Mid tibia whitish at the base, the remainder yellow with yellow hairs. Hind tibia white to yellowish at the base, the remainder brown with dark hairs. Fore tarsus black. Mid and hind tarsi with first tarsomere white, the remainder light brown. Anterior area of wing including pterostigma yellowish; a distinct dark brown apical patch extending in less intensity along the posterior margin to anal lobe. No appendix at fork. Halter yellow. Abdomen yellow, dorsally with yellow hairs in the middle and sides of tergites 1–5, black hairs in the remainder. Dorsolateral of tergites 4–7 with black patches, sometimes on the tergite 3, rarely absent on tergite 4. Sternites 1–5 yellow with yellow hairs, 6–7 black with black hairs.



Figures 1 – 4. *Diachlorus tenuimaculatus* n. sp. 1. holotype, habitus dorsal; 2. paratype, habitus lateral; 3. holotype, frontal view; 4. paratype, head lateral.

**Male.** Unknown.

**Type material.** All specimens previously stored in a tube with 70% ethanol and then pinned. Holotype female: PERU, *Cusco (sic)*, Quincemil, 633 m [elevation], 13°13'03"S; 70°43'40"W, 23–31.viii.2012, malaise [trap], J.A. Rafael, R.R. Cavichioli & D.M. Takiya [cols.] (MUSM); paratype females: same data as holotype (3 CEUFT, 10 INPA, 3 MPEG, 10 MUSM, 3 MZUSP); *Madre de Dios*, Mazuko, 382 m [elevation], 13°02'51"S; 70°20'46"W, 18–22.viii.2012, malaise [trap], R.R. Cavichioli, J.A. Rafael, A.P.M. Santos & D.M. Takiya [cols.] (5 INPA; 5 MUSM); *Cusco (sic)*, Quincemil, 874 m [elevation], 13°20'10"S; 70°50'57"W, viii. 2012, malaise, J.A. Rafael, A.P.M. Santos & D.M. Takiya (1 INPA).

**Etymology.** From Latin, *tenuis* = dilute, attenuated; *macula* = spot.

**Discussion.** Similar to *D. curvipes* (Fabricius) by the resemblance of scutum pattern, frons and legs, but the anterolateral spot in the scutum is somewhat conspicuous in the new species, absent in *D. curvipes*. Furthermore, the wing posterior border and the sides of tergites 4–7 are dark. It is similar to *D. heppneri* Wilkerson & Fairchild and *D. nuneztovari* Fairchild & Ortiz in the darkening of the wing posterior border, but both species do

not have the median yellow stripe on the scutum nor the dorsolateral black spots on the abdomen. *D. tenuimaculatus* n. sp. is somewhat similar to *D. varipes* (Rondani) in the darkening of the wing posterior border, but is readily differentiated by the wider frons, frontal index = 4, whereas in *D. varipes* = 7.

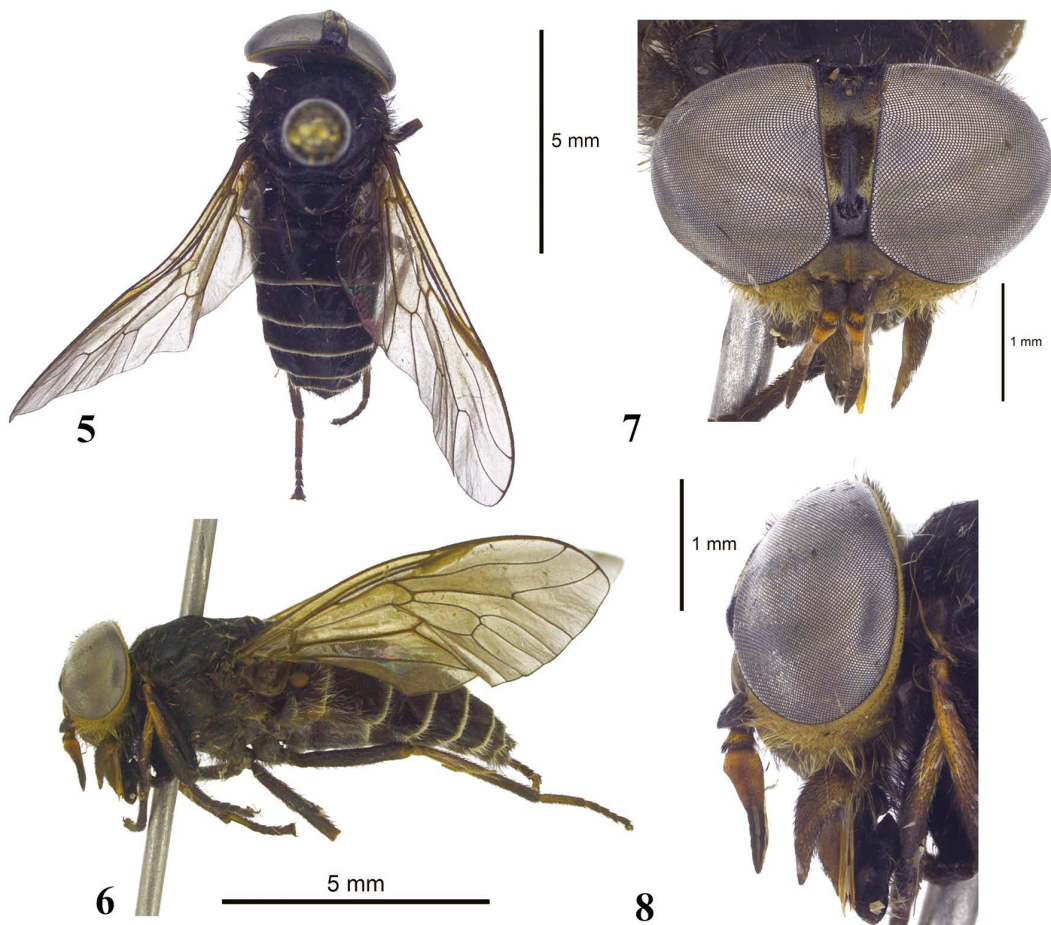
**Distribution.** Eastern Peru.

*Stenotabanus (Stenotabanus) carrascoi* n. sp.

(Figs. 5–8)

A black species, including notopleuron and pleuron. Frons divergent above. Abdominal segments 2 – 6 with whitish hind margin. Wing fumose with anterior region yellowish. Tibiae obscurely bicolored.

**Female.** Length of holotype 8.4 mm (variation of paratypes 7.5 – 9.7), wing 8.2 mm. Eye with two transverse bands. Frons whitish-gray pruinose, divergent above (divergence index = 1.7; frontal index = 4.4). Basal callus dark-brown to black, higher than wide. Median callus slender with lateral dark-brown to black patch of pruinose. Vertex shiny with vestiges of ocelli. Subcallus yellowish-brown pruinose. Subantennal band weak.



Figures 5–8. *Stenotabanus carrascoi* n. sp. 5. holotype, habitus dorsal; 6. holotype, habitus lateral; 7. holotype, frontal view; 8) holotype, head lateral.



Parafacial, clypeus and gena pale yellowish-brown generally with pale hairs; but there may be dark hairs near the antenna base as well as in the clypeus and gena. Antennal scape dark-brown, subshining, with black hairs. Pedicel and postpedicel orange-brown, style dark-brown. Palpus brown to dark-brown with black hairs. Proboscis black, membranous.

Scutum, scutellum and notopleuron black with black hairs. There are vestiges of greenish shining recumbent scale-like hairs (appear to have been removed by fixation in alcohol). Pleuron dark-brown pruinose with black hairs, except propleuron grayish pruinose and katatergite with a patch of pale hairs. Coxae and femora dark-brown to black with black hairs, the former may have gray pruinescence. Tibiae obscurely bicolored, brown "orange" with white hairs basally, dark-brown with black hairs apically, in the following proportion (pale/dark): fore tibia  $\frac{1}{2}$ , mid tibia  $\frac{1}{2}$  to  $\frac{3}{4}$ , hind tibia  $\frac{3}{4}$ . Sometimes the pale or the dark area is more extensive. Fore tarsus dark-brown, mid and hind tarsi brown, the remaining black. Wing fumose, bc, c, br, stem vein cell and pterostigma yellowish. Veins dark-brown. Halter brown. Abdomen black with hind margin of segments 2–6 whitish. The pilosity is predominantly black, except in the whitish margins, lateral of tergites 1–6 and sternites 1 and 2, with white hairs. Tergites 1 and 2 with gray-brown pruinescence, better visible from behind. Sternites 1 and 2 with grayish pruinescence.

**Male.** Unknown.

**Type material.** All specimens previously stored in a tube with 70% ethanol and then pinned. Holotype female: PERU, *Cusco (sic)*, Quincemil, 633 m [elevation], 13°13'03"S; 70°43'40"W, 23–31.viii.2012, malaise [trap], J.A. Rafael, R.R. Cavichioli & D.M. Takiya [cols.] (MUSM); paratype females: same data as holotype (3 CEUFT, 09 INPA, 3 MPEG, 10 MUSM, 3 MZUSP); idem, 874 m [elevation], 13°20'10"S; 70°50'57"W, viii. 2012, malaise, J.A. Rafael, A.P.M. Santos & D.M. Takiya (1 INPA).

**Etymology.** The name is an honor to Francisco Carrasco, Peruvian zoologist, who published in 1972 a list with 163 species for the country.

**Discussion.** Similar to *S. chaineyi* n. sp., described here, but has darker antenna, obscurely bicolored legs (tibiae), fumose wing and yellowish costal cell.

**Distribution.** Eastern Peru.

### *Stenotabanus (Stenotabanus) chaineyi* n. sp.

(Figs. 9–12)

*Stenotabanus* ?variant of *incipiens*; Chainey *et al.*, 1999: 90 (in key): 105 (discussion under *Stenotabanus incipiens*)

A distinct black species, including palpus, notopleuron and pleuron. Abdominal segments 2–6 with pale hind margins. Tibiae bicolored.

**Female.** Length of holotype 9 mm (variation of pa-

ratypes 7.6 – 9), wing. 8.8 mm. Eye with two transverse bands. Frons yellowish gray pruinose, wide and slightly divergent above (frontal index = 3.9; divergence index = 1.4). Basal callus black-brown, median callus black with black-brown lateral pruinescence, visible from below view. A conspicuous tubercle at vertex, but no ocelli. Subcallus with black-brown integument and yellowish gray pruinescence. Subantennal band brown to dark-brown. Clypeus, parafacial and gena yellowish gray pruinose with black hairs, sometimes pale hairs on parafacial and partially on gena. Antenna with scape and pedicel light brown to orange-brown with black hairs, slightly subshining, flagellum orange. Palpus black with black hairs. Proboscis black, membranous.

Scutum black, brownish gray anteriorly, with recumbent shining greenish hairs, scales-hair. Scutellum black with same scales, but whiter. Notopleuron black with erect black hairs and dorsal light scales. All pleuron dark-brown to black with dark-brown gray pruinescence and black hairs. All legs black with black hairs, except by bicolored tibiae, white at base in proportions: fore tibia  $\frac{1}{3}$ , mid tibia  $\frac{1}{2}$ , hind tibia  $\frac{2}{3}$ . All tarsi black. Wing hyaline with brown to dark-brown veins; the unique areas tinged are bc cell and stem vein cell brown, and pterostigma yellowish. Halter dark-brown. Abdomen mostly black with pale hind margin of segments 2–6 or 7. Hairs color same as the background, except sternite 2 where there are black and white hairs. Sternites 1 and 2 with bluish-gray pruinescence.

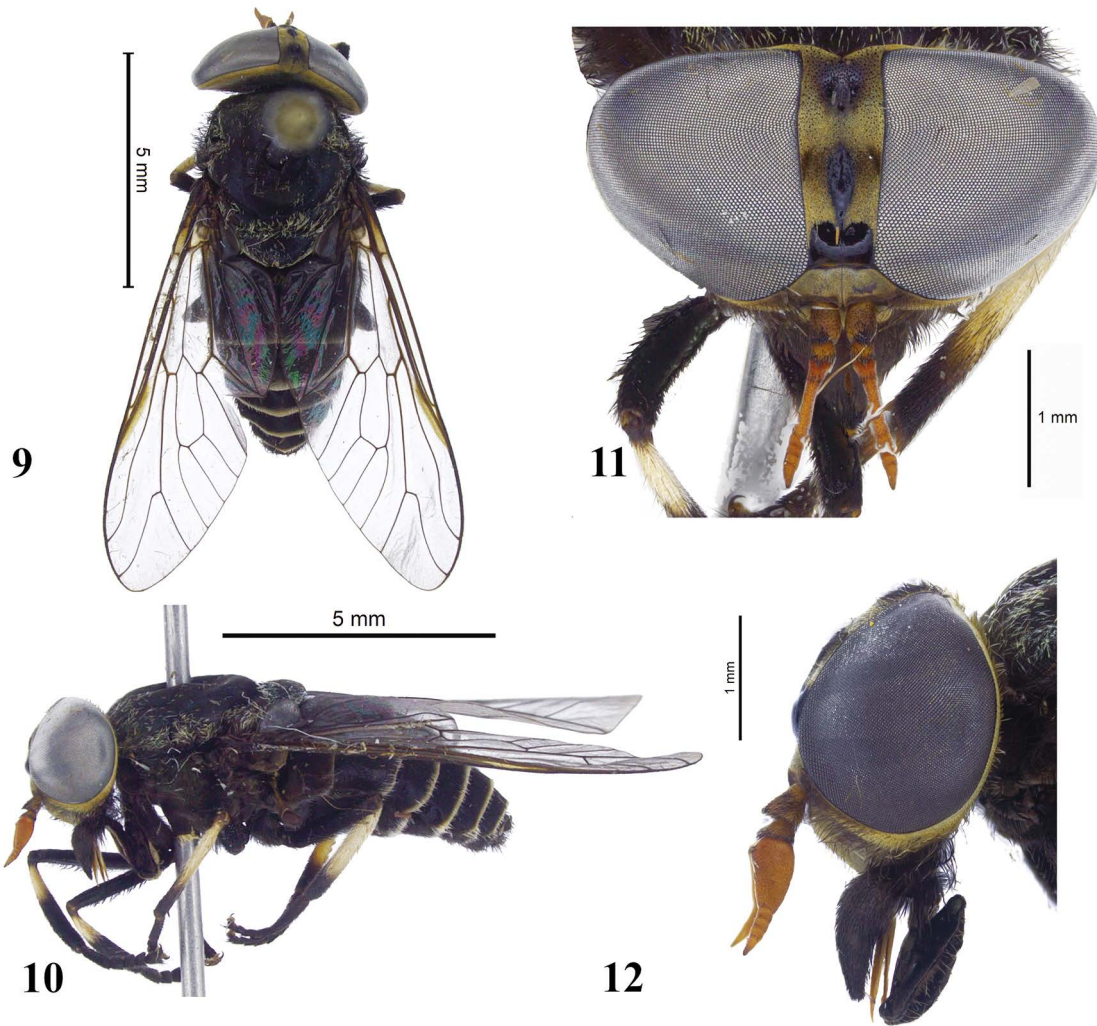
**Male.** Unknown.

**Type material.** All specimens previously stored in a tube with 70% ethanol and then pinned. Holotype female: PERU, *Madre de Dios*, Mazuco, 382 m [elevation], 13°02'51"S; 70°20'46"W, 18–22.viii.2012, malaise [trap], R.R. Cavichioli, J.A. Rafael, A.P.M. Santos & D.M. Takiya [cols.] (MUSM); paratype females: same data as holotype (2 CEUFT, 6 INPA, 2 MPEG, 4 MUSM, 2 MZUSP); *Cusco (sic)*, Quincemil, 633 m [elevation], 13°13'03"S; 70°43'40"W, 23–31.viii.2012, malaise, J.A. Rafael, R.R. Cavichioli & D.M. Takiya (1 CEUFT, 2 INPA, 1 MPEG, 2 MUSM, 1 MZUSP).

**Etymology.** The name is in honor of John E. Chainey, retired researcher at the Natural History Museum (London), who first noticed the possibility of a new taxon.

**Discussion.** It resembles *Stenotabanus incipiens*, but it is easily separable by having black palpus and pleuron; tergites 1–2 lacking the bluish-gray lateral pruinescence; and tergites 2–6 with narrower, pale hind margins that are less expanded medially. Chainey *et al.* (1999: 105) examined specimens from Peru and Bolivia of the species described here, but hesitated to describe it, however, they made comparisons and commented on variations such as the presence of light hairs on the clypeus and parafacial.

**Distribution.** Eastern Peru and Bolivia.



Figures 9 – 12. *Stenotabanus chaineyi* n. sp. 9. holotype, habitus dorsal; 10. holotype, habitus lateral; 11. holotype, frontal view; 12. holotype, head lateral.

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ALH: Ideas; formulation of overarching research goals and aims, described the new species and updated the list of species; TKK: who accepted the challenge, jointly described the three new species, and assisted in updating the species list.

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**Appendix 1.** Species of Tabanidae captured by Malaise traps in Peru, August 2012.

Collection sites:

- 1 – Madre de Dios, Mazuko, elevation 382 m, 13°02'51"S; 70°20'46"W
- 2 – Cuzco, Quincemil, elevation 633 m, 13°13'03"S; 70°43'40"W
- 3 – Cuzco, Quincemil, elevation 874 m, 13°20'10"S; 70°50'57"W
- 4 – Cuzco, Quincemil, elevation 1000 m, 13°21'18"S; 70°53'22"W

Species	Site(s)	sex (Collection)
<i>Esenbeckia testaceiventris</i> (Macquart), 1848	2	1 female (INPA)
<i>Scione</i> spp.	2,3,4	6 females (INPA, MUSM)
<i>Betrequia ocellata</i> Oldroyd, 1970	1	1 male (INPA)
<i>Chrysops leucospilus</i> Wiedemann, 1828	1,2,4	4 females (INPA, MUSM)
<i>Chrysops varians</i> Wiedemann, 1828	1,2,3	5 females (INPA, MUSM)
<i>Bolbodimyia celeroides</i> Stone, 1954	1,2	3 females (INPA, MUSM)
<i>Bolbodimyia desepta</i> Enderlein, 1925	2	1 female (INPA)
<i>Bolbodimyia nigra</i> Stone, 1934	2,3	2 females (INPA, MUSM)
<i>Catachlorops striatus</i> Burger, 1999	1,2	7 females (INPA, MUSM)
<i>Catachlorops vespertinus</i> (Bequaert & Renjifo-Salcedo), 1946	1,2,3,4	56 females (CEUFT, INPA, MUSM)
<i>Chlorotabanus inanis</i> (Fabricius), 1787	2	1 female (INPA)
<i>Dasychela ?fulvicornis</i> (Kröber), 1931	2	2 females (INPA, MUSM)
<i>Diachlorus tenuimaculatus</i> n. sp.	1,2,3	Holotype female (MUSM), paratypes female (3 CEUFT, 16 INPA, 3 MPEG, 15 MUSM, 3 MZUSP)
<i>Diachlorus trevori</i> Wilkerson & Fairchild, 1982	1	27 males, 5 females (INPA, MUSM)
<i>Diachlorus xynus</i> Fairchild, 1972	2	1 female (INPA)
<i>Dichelacera submarginata</i> Lutz, 1915	1,2,3,4	32 females (CEUFT, INPA, MUSM)
<i>Di cladocera</i> spp.	1,2,3,4	13 females (INPA, MUSM)
<i>Himantostylus intermedius</i> Lutz, 1913	1,3	2 females (INPA, MUSM)
<i>Leucotabanus weyrauchi</i> Fairchild, 1951	1,2,3	3 males and 1 female (INPA, MUSM)
<i>Phaeotabanus nigriflavus</i> (Kröber), 1930	1	2 females (INPA, MUSM)
<i>Phaeotabanus phaeopterus</i> Fairchild, 1964	1,2,3	7 males, 78 females (INPA, MUSM)
<i>Philipotabanus opimus</i> Fairchild, 1975	1,2	80 females (INPA, MUSM)
<i>Poeciloderas quadripunctatus</i> (Fabricius), 1805	1,2,3	1 male, 42 females (INPA, MUSM)
<i>Stenotabanus albilinearis</i> Philip, 1960	1	7 females (CEUFT, INPA, MUSM)
<i>Stenotabanus albiscutellatus</i> Chainey, 1999	1	4 females (INPA, MUSM)
<i>Stenotabanus carrascoi</i> n. sp.	2,3	Holotype female (MUSM), paratypes female (3 CEUFT, 10 INPA, 3 MPEG, 10 MUSM, 3 MZUSP)
<i>Stenotabanus chaineyi</i> n. sp.	1,2	Holotype female (MUSM), paratypes female (3 CEUFT, 8 INPA, 3 MPEG, 6 MUSM, 3 MZUSP)
<i>Stenotabanus nigriscapus</i> Chainey, 1999	1,2,3	82 females (CEUFT, INPA, MUSM)
<i>Stenotabanus obscurus</i> Kröber, 1929	2	1 male, 15 females (CEUFT, INPA, MUSM)
<i>Stypommisa apicalis</i> Fairchild & Wilkerson, 1986	1	2 females (INPA, MUSM)
<i>Stypommisa captiroptera</i> (Kröber), 1930	1	3 females (INPA, MUSM)
<i>Stypommisa glandicolor</i> (Lutz), 1912	1	9 females (INPA, MUSM)
<i>Stypommisa furva</i> (Hine), 1920	2,4	47 females (CEUFT, INPA, MUSM)
<i>Stypommisa kroeberi</i> Fairchild & Wilkerson, 1986	1,2,4	12 females (CEUFT, INPA, MUSM)
<i>Stypommisa spilota</i> Fairchild & Wilkerson, 1986	1,2,3,4	1 male, 60 females (CEUFT, INPA, MUSM)
<i>Stypommisa venosa</i> (Bigot), 1892	2,4	3 females (INPA, MUSM)
<i>Tabanus hirtitibia</i> Walker, 1850	1,2	7 females (INPA, MUSM)
<i>Tabanus occidentalis</i> Linnaeus, 1758	1,2	24 females (INPA, MUSM)
<i>Tabanus sextriangulus</i> Gorayeb & Rafael, 1984	1	2 females (INPA, MUSM)
<i>Tabanus weyrauchi</i> (Barretto), 1949	1	1 female (INPA)

**Appendix 2.** Updated list of species of Tabanidae of Peru**Pangoniinae** (53 spp.)**Pangoniini** (21 spp.)

1. *Esenbeckia (Esenbeckia) bassleri* Wilkerson & Fairchild, 1983
2. *Esenbeckia (Esenbeckia) bitriangulata* Lutz & Castro, 1935
3. *Esenbeckia (Esenbeckia) cisandeanana* Wilkerson & Fairchild, 1983
4. *Esenbeckia (Esenbeckia) gracilis* Kröber, 1931
5. *Esenbeckia (Esenbeckia) melanogaster* Lutz & Castro, 1935
6. *Esenbeckia (Esenbeckia) nigriventris* Kröber, 1931
7. *Esenbeckia (Esenbeckia) osornoi* Fairchild, 1942
8. *Esenbeckia (Esenbeckia) pechumani* Wilkerson & Fairchild, 1983
9. *Esenbeckia (Esenbeckia) perspicua* Wilkerson & Fairchild, 1983
10. *Esenbeckia (Esenbeckia) peruviana* Burger, 1999
11. *Esenbeckia (Esenbeckia) testaceiventris* (Macquart), 1848
12. *Esenbeckia (Esenbeckia) tigrina* Wilkerson, 1979
13. *Esenbeckia (Esenbeckia) translucens* (Macquart), 1846
14. *Esenbeckia (Esenbeckia) vulpes* (Wiedemann), 1828
15. *Esenbeckia (Esenbeckia) wygodzinskyi* Wilkerson & Fairchild, 1983
16. *Esenbeckia (Esenbeckia) xanthoskela* Wilkerson & Fairchild, 1983
17. *Esenbeckia (Proboscoides) arcuata* (Williston), 1895
18. *Esenbeckia (Proboscoides) fairchildi* (Philip), 1943
19. *Esenbeckia (Proboscoides) rostrum* (Philip), 1943
20. *Esenbeckia (Proboscoides) schlingerii* Philip, 1960
21. *Esenbeckia (Proboscoides) suturalis* (Rondani), 1848

**Scionini** (32 spp.)

22. *Fidena (Fidena) atripes* (Röder), 1886
23. *Fidena (Fidena) brevistria* (Lutz), 1909
24. *Fidena (Fidena) eriomerooides* (Lutz), 1909
25. *Fidena (Fidena) freemani* Barretto, 1957
26. *Fidena (Fidena) kroeberi* Fairchild, 1971
27. *Fidena (Fidena) nigripes* (Röder), 1886
28. *Fidena (Fidena) ochrapogon* Wilkerson, 1979
29. *Fidena (Fidena) rhinophora* (Bellardi), 1859
30. *Fidena (Fidena) zonalis* Kröber, 1931
31. *Fidena (Laphriomyia) silvatica* (Brèthes), 1920
32. *Oscia aureonigra* Kröber, 1931
33. *Oscia rubriventris* Kröber, 1930
34. *Pityocera (Elaphella) cervus* (Wiedemann), 1828
35. *Scione acris* Philip, 1958
36. *Scione albohirta* Kröber, 1930
37. *Scione aurulans lurida* Enderlein, 1925
38. *Scione brevistriga* Enderlein, 1925
39. *Scione cingulata* (Enderlein), 1925
40. *Scione claripennis* Ricardo, 1900
41. *Scione crassa* Szilády, 1926
42. *Scione equivexans* Wilkerson, 1979
43. *Scione flavohirta* Ricardo, 1902
44. *Scione fulva* Ricardo, 1902
45. *Scione fumipennis* Kröber, 1930
46. *Scione fusca* Ricardo, 1900

47. *Scione grandis* Philip, 1943
48. *Scione huancabambae* Kröber, 1930
49. *Scione limbativena* Enderlein, 1925
50. *Scione longirostris* Brèthes, 1920
51. *Scione obscurefemorata* Kröber, 1930
52. *Scione rufescens* (Ricardo), 1900
53. *Scione strigata* (Enderlein), 1925

**Chrysopsinae** (14 spp.)**Chrysopsini** (13 spp.)

54. *Chrysops boliviensis* Kröber, 1926
55. *Chrysops bulbicornis* Lutz, 1911
56. *Chrysops calogaster* Schiner, 1868
57. *Chrysops ecuadorensis* Lutz, 1909
58. *Chrysops incisus* Macquart, 1846
59. *Chrysops laetus* Fabricius, 1805
60. *Chrysops leucospilus* Wiedemann, 1828
61. *Chrysops molestus* Wiedemann, 1828
62. *Chrysops peruvianus* Kröber, 1925
63. *Chrysops rossi* Philip, 1960
64. *Chrysops varians* Wiedemann, 1828
65. *Chrysops variegatus* (De Geer), 1776
66. *Chrysops weberi* Bequaert, 1946

**Rhinomyzini** (1 sp.)

67. *Betrequia ocellata* Oldroyd, 1970

**Tabaninae** (166 spp.)**Diachlorini** (124 spp.)

68. *Acanthocera (Acanthocera) fairchildi* Henriques & Rafael, 1992
69. *Acanthocera (Acanthocera) gorayebi* Henriques & Rafael, 1992
70. *Acanthocera (Acanthocera) marginalis* Walker, 1854
71. *Acanthocera (Querbetia) chaineyi* Fairchild & Burger, 1994
72. *Acanthocera (Querbetia) inopinata* (Fairchild), 1972
73. *Agelanius antenninus* (Philip), 1969
74. *Bolbodimyia bicolor* Bigot, 1892
75. *Bolbodimyia celerooides* Stone, 1954
76. *Bolbodimyia desepta* Enderlein, 1925
77. *Bolbodimyia nigra* Stone, 1934
78. *Catachlorops (Amphichlorops) striatus* Burger, 1999
79. *Catachlorops (Amphichlorops) vespertinus* (Bequaert & Renjifo-Salcedo), 1946
80. *Catachlorops (Catachlorops) fumipennis* Kröber, 1931
81. *Catachlorops (Catachlorops) halteratus* Kröber, 1931
82. *Catachlorops (Hadrochlorops) scutellatus* (Macquart), 1838
83. *Catachlorops (Psalidia) rubiginosus* (Summers), 1911
84. *Chlorotabanus inanis* (Fabricius), 1787
85. *Chlorotabanus leucochlorus* Fairchild, 1961
86. *Chlorotabanus leuconotus* Krolow & Henriques, 2010
87. *Chlorotabanus parviceps* (Kröber), 1934
88. *Cryptotylus unicolor* (Wiedemann), 1828
89. *Dasybasis adornata* (Kröber), 1934
90. *Dasybasis albotibialis* (Kröber), 1931
91. *Dasybasis barbata* Coscarón & Philip, 1967

92. *Dasybasis bulbiscapens* Coscarón & Philip, 1967  
 93. *Dasybasis chilensis* (Macquart), 1838  
 94. *Dasybasis fairchildi* Coscarón & Philip, 1967  
 95. *Dasybasis frequens* (Kröber), 1934  
 96. *Dasybasis geminata* Coscarón & Philip, 1967  
 97. *Dasybasis inata* Coscarón & Philip, 1967  
 98. *Dasybasis kroeberi* Coscarón & Philip, 1967  
 99. *Dasybasis limbativena* (Kröber), 1931  
 100. *Dasybasis montium* (Surcouf), 1919  
 101. *Dasybasis neogrisescens* (Kröber), 1934  
 102. *Dasybasis penai* Coscarón & Philip, 1967  
 103. *Dasybasis punensis* (Hine), 1920  
 104. *Dasybasis schineri* (Kröber), 1931  
 105. *Dasybasis schnusei* (Kröber), 1931  
 106. *Dasychela (Dasychela) amazonensis* (Barretto), 1946  
 107. *Dasychela (Dasychela) fulvicornis* (Kröber), 1931  
 108. *Dasychela (Dasychela) inca* (Philip), 1960  
 109. *Dasychela (Dasychela) peruviana* (Bigot), 1892  
 110. *Dasychela (Triceratomyia) biramula* Fairchild, 1958  
 111. *Dasychela (Triceratomyia) macintyreii* (Bequaert), 1937  
 112. *Diachlorus anduzei* Stone, 1944  
 113. *Diachlorus bicinctus* (Fabricius), 1805  
 114. *Diachlorus bimaculatus* (Wiedemann), 1828  
 115. *Diachlorus curvipes* (Fabricius), 1805  
 116. *Diachlorus fuscistigma* Lutz, 1913  
 117. *Diachlorus heppneri* Wilkerson & Fairchild, 1982  
 118. *Diachlorus nuneztovari* Fairchild & Ortiz, 1955  
 119. *Diachlorus pechumani* Fairchild, 1972  
 120. *Diachlorus scutellatus* (Macquart), 1838  
 121. *Diachlorus tenuimaculatus* n. sp.  
 122. *Diachlorus trevori* Wilkerson & Fairchild, 1982  
 123. *Diachlorus xynus* Fairchild, 1972  
 124. *Dichelacera (Desmatochelacera) albitibialis* Burger, 1999  
 125. *Dichelacera (Dichelacera) cervicornis* (Fabricius), 1805  
 126. *Dichelacera (Dichelacera) marginata* Macquart, 1847  
 127. *Dichelacera (Dichelacera) rubrofemorata* Burger, 1999  
 128. *Dichelacera (Dichelacera) submarginata* Lutz, 1915  
 129. *Dichelacera (Dichelacera) villavoensis* Fairchild & Philip, 1960  
 130. *Dichelacera (Idiochelacera) subcallosa* Fairchild & Philip, 1960  
 131. *Di cladocera argentomacula* Wilkerson, 1979  
 132. *Di cladocera bellicosa* (Brèthes), 1910  
 133. *Di cladocera castanea* (Barretto), 1949  
 134. *Di cladocera clara* (Schiner), 1868  
 135. *Di cladocera fairchildi* Goodwin, 1999  
 136. *Di cladocera griseipennis* Kröber, 1931  
 137. *Di cladocera hoppi* Enderlein, 1927  
 138. *Di cladocera macula* (Macquart), 1846  
 139. *Di cladocera maculistigma* Enderlein, 1925  
 140. *Di cladocera minos* (Schiner), 1868  
 141. *Di cladocera neosubmacula* Kröber, 1931  
 142. *Di cladocera nigrocoerulea* (Rondani), 1850  
 143. *Di cladocera nova* Kröber, 1931  
 144. *Eutabanus pictus* Kröber, 1930  
 145. *Haematopotina pechumani* (Coscarón & Philip), 1967  
 146. *Himantostylus intermedius* Lutz, 1913  
 147. *Lepiselaga crassipes* (Fabricius), 1805  
 148. *Leucotabanus albovarius* (Walker), 1854  
 149. *Leucotabanus exaestuans* (Linnaeus), 1758  
 150. *Leucotabanus flavinotum* (Kröber), 1934  
 151. *Leucotabanus weyrauchi* Fairchild, 1951  
 152. *Montismyia lauta* (Hine), 1920  
 153. *Phaeotabanus aphanopterus* (Wiedemann), 1828  
 154. *Phaeotabanus cajennensis* (Fabricius), 1787  
 155. *Phaeotabanus fervens* (Linnaeus), 1758  
 156. *Phaeotabanus innotescens* (Walker), 1854  
 157. *Phaeotabanus nigriflavus* (Kröber), 1930  
 158. *Phaeotabanus phaeopterus* Fairchild, 1964  
 159. *Phaeotabanus serenus* (Kröber), 1931  
 160. *Philipotabanus (Mimotabanus) fucosus* Fairchild, 1958  
 161. *Philipotabanus (Mimotabanus) opimus* Fairchild, 1975  
 162. *Philipotabanus (Mimotabanus) phalaropygus* Fairchild, 1964  
 163. *Philipotabanus (Philipotabanus) reticulatus* (Kröber), 1930  
 164. *Philipotabanus (Philipotabanus) stigmatalis* (Kröber), 1931  
 165. *Philipotabanus (Philipotabanus) tenuifasciatus* (Kröber), 1930  
 166. *Spilotabanus multiguttatus* (Kröber), 1930  
 167. *Stenotabanus (Stenotabanus) albilinearis* Philip, 1960  
 168. *Stenotabanus (Stenotabanus) albiscutellatus* Chainey, 1999  
 169. *Stenotabanus (Stenotabanus) bruesi* (Hine), 1920  
 170. *Stenotabanus (Stenotabanus) carrascoi* n. sp.  
 171. *Stenotabanus (Stenotabanus) chaineyi* n. sp.  
 172. *Stenotabanus (Stenotabanus) incipiens* (Walker), 1860  
 173. *Stenotabanus (Stenotabanus) nigriscapus* Chainey & Hall, 1999  
 174. *Stenotabanus (Stenotabanus) obscurus* Kröber, 1929  
 175. *Stenotabanus (Stenotabanus) penai* Chainey, 1999  
 176. *Stenotabanus (Stenotabanus) peruviansis* Kröber, 1929  
 177. *Stibasoma flaviventris* (Macquart), 1848  
 178. *Stypommisa anoriensis* Fairchild & Wilkerson, 1986  
 179. *Stypommisa apicalis* Fairchild & Wilkerson, 1986  
 180. *Stypommisa captiroptera* (Kröber), 1930  
 181. *Stypommisa flavescens* (Kröber), 1930  
 182. *Stypommisa furva* (Hine), 1920  
 183. *Stypommisa glandicolor* (Lutz), 1912  
 184. *Stypommisa hypographa* (Kröber), 1930  
 185. *Stypommisa hypographa neofurva* Philip, 1969  
 186. *Stypommisa kroeberi* Fairchild & Wilkerson, 1986  
 187. *Stypommisa modica* (Hine), 1920  
 188. *Stypommisa pequeniensis* (Fairchild), 1942  
 189. *Stypommisa prunicolor* (Lutz), 1912  
 190. *Stypommisa spilota* Fairchild & Wilkerson, 1986  
 191. *Stypommisa venosa* (Bigot), 1892
- Tabanini (42 spp.)**
192. *Phorcotabanus cinereus* (Wiedemann), 1821

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193. *Poeciloderas caesiomaculatus* (Kröber), 1931  
194. *Poeciloderas ornatipennis* (Kröber), 1934  
195. *Poeciloderas quadripunctatus* (Fabricius), 1805  
196. *Tabanus angustifrons* Macquart, 1848  
197. *Tabanus antarcticus* Linnaeus, 1758  
198. *Tabanus argentivittatus* Fairchild, 1976  
199. *Tabanus argentivittatus huallagensis* Fairchild, 1976  
200. *Tabanus basilaris* Kröber, 1931  
201. *Tabanus boliviensis* (Kröber), 1930  
202. *Tabanus callosus* Macquart, 1848  
203. *Tabanus colombensis* Macquart, 1846  
204. *Tabanus discifer* Walker, 1850  
205. *Tabanus fortis* Fairchild, 1961  
206. *Tabanus fumomarginatus* Hine, 1920  
207. *Tabanus glaucus* Wiedemann, 1819  
208. *Tabanus guyanensis* Macquart, 1846  
209. *Tabanus hirtitibia* Walker, 1850  
210. *Tabanus importunus* Wiedemann, 1828  
211. *Tabanus isis* Fairchild, 1976  
212. *Tabanus macquarti* Schiner, 1868  
213. *Tabanus nebulosus* De Geer, 1776  
214. *Tabanus nematocallus* Fairchild, 1984  
215. *Tabanus noncallosus* Carmo & Henriques, 2019  
216. *Tabanus occidentalis* Linnaeus, 1758  
217. *Tabanus pellucidus* Fabricius, 1805  
218. *Tabanus penai* Philip, 1967  
219. *Tabanus piceiventris* Rondani, 1848  
220. *Tabanus picicallosus* Fairchild, 1951  
221. *Tabanus pungens* Wiedemann, 1828  
222. *Tabanus restrepoensis* Fairchild, 1942  
223. *Tabanus rubripes* Macquart, 1838  
224. *Tabanus sannio* Fairchild, 1956  
225. *Tabanus secundus* Walker, 1848  
226. *Tabanus sextriangulus* Gorayeb & Rafael, 1984  
227. *Tabanus sorbillans* Wiedemann, 1828  
228. *Tabanus trivittatus* Fabricius, 1805  
229. *Tabanus weyrauchi* (Barretto), 1949  
230. *Tabanus wilkersoni* Fairchild, 1983  
231. *Tabanus xanthogaster* Philippi, 1865  
232. *Tabanus xerodes* Philip, 1967  
233. *Tabanus xuthopogon* Fairchild, 1984
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